



# Dietary fiber for the prevention of childhood obesity: a focus on the involvement of the gut microbiota

Zhongmin Yang, Mingyue Yang, Edward C. Deehan, Chenxi Cai, Karen L. Madsen, Eytan Wine, Guiling Li, Jian Li, Jingwen Liu & Zhengxiao Zhang

To cite this article: Zhongmin Yang, Mingyue Yang, Edward C. Deehan, Chenxi Cai, Karen L. Madsen, Eytan Wine, Guiling Li, Jian Li, Jingwen Liu & Zhengxiao Zhang (2024) Dietary fiber for the prevention of childhood obesity: a focus on the involvement of the gut microbiota, Gut Microbes, 16:1, 2387796, DOI: [10.1080/19490976.2024.2387796](https://doi.org/10.1080/19490976.2024.2387796)

To link to this article: <https://doi.org/10.1080/19490976.2024.2387796>



© 2024 The Author(s). Published with license by Taylor & Francis Group, LLC.



[View supplementary material](#)



Published online: 20 Aug 2024.



[Submit your article to this journal](#)













[View related articles](#)



[View Crossmark data](#)

# Dietary fiber for the prevention of childhood obesity: a focus on the involvement of the gut microbiota

Zhongmin Yang <sup>a</sup>, Mingyue Yang <sup>a</sup>, Edward C. Deehan <sup>b,c</sup>, Chenxi Cai <sup>d</sup>, Karen L. Madsen <sup>e</sup>, Eytan Wine <sup>f</sup>, Guiling Li <sup>a,g</sup>, Jian Li <sup>a,g</sup>, Jingwen Liu <sup>a</sup>, and Zhengxiao Zhang <sup>a,g</sup>

<sup>a</sup>College of Ocean Food and Biological Engineering, Jimei University, Xiamen, Fujian, China; <sup>b</sup>Department of Food Science and Technology, University of Nebraska, Lincoln, NE, USA; <sup>c</sup>Nebraska Food for Health Center, University of Nebraska, Lincoln, NE, USA; <sup>d</sup>School of Public Health, Xiamen University, Xiamen, Fujian, China; <sup>e</sup>Division of Gastroenterology, Department of Medicine, University of Alberta, Edmonton, AB, Canada; <sup>f</sup>Division of Pediatric Gastroenterology, Departments of Pediatrics and Physiology, University of Alberta, Edmonton, AB, Canada; <sup>g</sup>Fujian Provincial Engineering Technology Research Center of Marine Functional Food, Xiamen, Fujian, China

## ABSTRACT

Given the worldwide epidemic of overweight and obesity among children, evidence-based dietary recommendations are fundamentally important for obesity prevention. Although the significance of the human gut microbiome in shaping the physiological effects of diet and obesity has been widely recognized, nutritional therapeutics for the mitigation of pediatric obesity globally are only just starting to leverage advancements in the nutritional microbiology field. In this review, we extracted data from PubMed, EMBASE, Scopus, Web of Science, Google Scholar, CNKI, Cochrane Library and Wiley online library that focuses on the characterization of gut microbiota (including bacteria, fungi, viruses, and archaea) in children with obesity. We further review host-microbe interactions as mechanisms mediating the physiological effects of dietary fibers and how fibers alter the gut microbiota in children with obesity. Contemporary nutritional recommendations for the prevention of pediatric obesity are also discussed from a gut microbiological perspective. Finally, we propose an experimental framework for integrating gut microbiota into nutritional interventions for children with obesity and provide recommendations for the design of future studies on precision nutrition for pediatric obesity.

## ARTICLE HISTORY

Received 27 February 2024  
Revised 29 June 2024  
Accepted 30 July 2024

## KEYWORDS


Dietary fiber; gut microbiota; childhood; obesity; precision nutrition


## 1. Introduction

The presence of childhood obesity is a prominent risk factor for the development of obesity-related comorbidities in adolescence, including increased risk of type 2 diabetes<sup>1</sup>, nonalcoholic fatty liver disease,<sup>2</sup> and cardiovascular disease.<sup>3</sup> Data released by the World Obesity Federation reported that the increase in the prevalence of obesity is expected to be steepest among children and adolescents aged 5–19 years, rising from 10% to 20% of the world's boys and from 8% to 18% of the world's girls between 2020 and 2035.<sup>4</sup> Recent research further suggests that having excess body weight in childhood and adolescence was not only found to adversely affect physical health later in life<sup>5</sup> but also negatively impact psychosocial wellbeing.<sup>6</sup> Nevertheless, current strategies for treating childhood obesity, such as lifestyle changes and pharmacotherapy, have very modest effectiveness,<sup>7</sup> concerning side effects,<sup>8</sup> or inconsistent outcomes.<sup>9</sup>

Identifying the causes of childhood obesity and targeting interventions is, therefore, important in the context of preventive health.

Various factors have been suggested as potential causes of obesity, including lifestyle choices and genetic factors,<sup>10</sup> with the gut microbiota emerging as an important factor contributing to the pathogenesis of obesity.<sup>11</sup> Due to their profound effects on the host, this highly complex group of microbes that reside along the human gastrointestinal tract is referred to as an individual's "second genome".<sup>12</sup> The gut microbiota is a community of more than 10<sup>13</sup> microorganisms that includes bacteria, fungi, viruses, and archaea, which live as a complex ecosystem with the human host.<sup>6,13</sup> There is accumulating evidence that the gut microbiota affects all aspects of energy homeostasis through mechanisms involving immune, hormonal and neural

**CONTACT** Jingwen Liu  [ljwsbch@163.com](mailto:ljwsbch@163.com); Zhengxiao Zhang  [zxzhang@jmu.edu.cn](mailto:zxzhang@jmu.edu.cn)  College of Ocean Food and Biological Engineering, Jimei University, 185 Yinjiang Road, Jimei District, Xiamen, Fujian 361021, China

 Supplemental data for this article can be accessed online at <https://doi.org/10.1080/19490976.2024.2387796>

© 2024 The Author(s). Published with license by Taylor & Francis Group, LLC.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0/>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited. The terms on which this article has been published allow the posting of the Accepted Manuscript in a repository by the author(s) or with their consent.

systems, and that gut dysbiosis, or an abnormal composition of gut microbial taxa might contribute to a disturbed host metabolism through effects on adipose tissue, muscle, and liver.<sup>14</sup>

Although growing evidence from animal models that the gut microbiota has a potential causal role in obesity and would be a promising target for precision therapeutics, there is still a lack of convincing evidence for a direct contribution of the gut microbiota to humans with obesity.<sup>15</sup> Whether gut microbial alteration exists in children with obesity is yet to be systematically evaluated.<sup>16,17</sup> Moreover, the gut microbiome is thought to stabilize and resemble that of an adult after the first 1 to 3 years of life,<sup>18</sup> while other research suggests that it continues to develop throughout adolescence and that gaining excess body weight during childhood is likely to play a role in further shaping the gut microbiota.<sup>19,20</sup> It is unclear what role the coexistence of gut cross-kingdom community interactions in the gut plays in childhood obesity. Therefore, identifying the gut microbiota characteristics that are conserved in children with obesity and also play a pathogenic role in the development of childhood obesity is a challenge in the field. This has important implications for the development of microbiome-targeted therapies for the prevention and treatment of childhood obesity.

Dietary substances, such as fibers and prebiotics, have been found to induce a positive impact on the children gut microbiota.<sup>21</sup> As a result, they hold potential for the management of obesity. Dietary fibers (DFs), carbohydrate polymers that are neither digested nor absorbed in the small intestine, are subject to microbial fermentation in the human colon and, thus, influence the composition of microbial communities as well as their metabolic activities.<sup>22</sup> Some DFs can also be classified as prebiotics,<sup>23</sup> or substrates selectively used by host microorganisms that confer a health benefit to humans.<sup>23</sup> However, currently it is not clear if the beneficial effects of increasing DF intake are due to microbial metabolism and the production of metabolites such as short-chain fatty acids (SCFAs) or due to a DF-altered intestinal transit time, changes in nutrient absorption, fecal bulking and binding of various metabolites, or modulation of the immune system.<sup>18,24</sup>

It is crucial to keep in mind that the physiological functions of DFs are highly structure-dependent. Subtle variations in the chemical structures of DFs affect their utilization by gut microbes, as bacteria have different abilities to cleave the linkages in the structure of these complex molecules to obtain simple sugars.<sup>25</sup> Simultaneously, recent reviews have revealed that intrinsic and isolated fibers have functional specificity as well.<sup>26</sup> However, there is insufficient literature evidence to support the targeted modulatory effects of DFs on the gut microbiota of children with obesity. Furthermore, adults are encouraged to consume at least 25–38 g of DFs per day, but there is currently a lack of data to support DF guidelines for children in most countries. As a result, more research is needed to thoroughly investigate the relationship between DFs and childhood obesity from a gut microbiota perspective.

In studies of children with obesity, examination of DF-microbiota-host interactions has been limited, despite recent increases in nutritional microbiology research in adults. In this perspective, we first systematically evaluated whether special microbial alterations exist in children with obesity, and then reviewed the effects of DFs and DFs-related patterns on the gut microbiota in children with obesity. We further apply this information to confront an ongoing debate in the field of childhood obesity research. Finally, we propose a microbiome-targeted research approach for evaluating treatments for the reduction of childhood obesity and outline an experimental framework for systematically incorporating the gut microbiome into future nutrition research.

## **2. Systematic review: characterization of the gut microbiota in children with obesity**

We conducted a systematic literature review in July 2022 (with an updated literature search in June 2024) to assess the pertinent gut microbiota traits of childhood obesity (see Box 1 for search strategy). Of the 63 studies that met the inclusion criteria, some of the gut microbes in children with obesity showed consistent trends (e.g., Firmicutes/Bacteroidetes (F/B) ratios, *Akkermansia muciniphila*, *Lactobacillus* spp., *Dialister*, etc). However, there is considerable heterogeneity among the findings of most published studies investigating gut microbiota

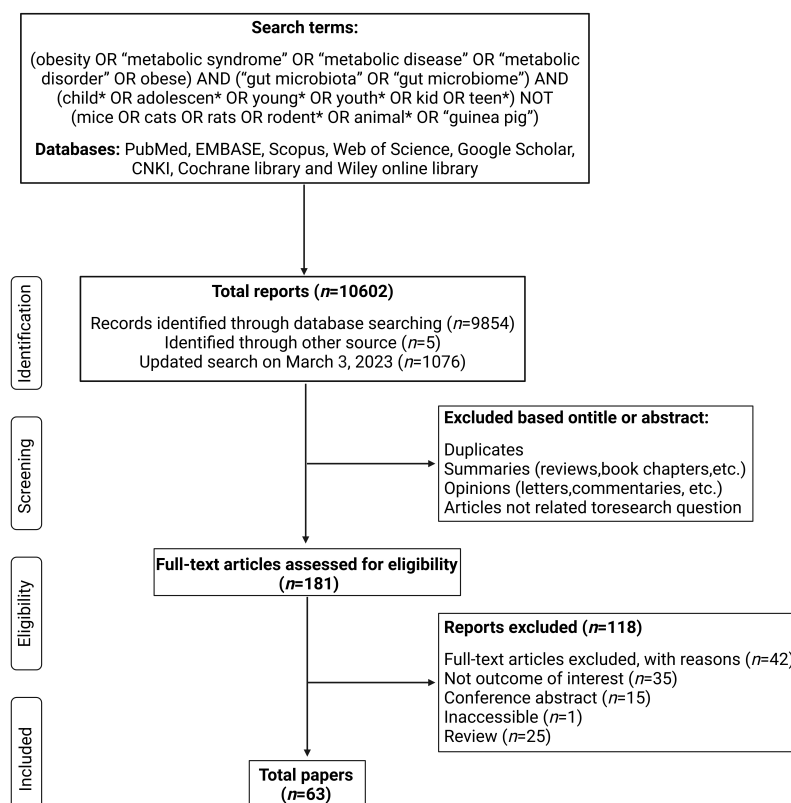
in childhood obesity. Understanding the compositional characteristics of the gut microbiota has the potential to help determine their role in the onset or development of obesity in the host, laying the groundwork for microbiome-based therapies for childhood obesity to improve the quality of life.

(1 species) in children with obesity in agreement with the indicated relationship with adiposity. The proceeding detailed analysis is grouped by gut bacterial diversity and compositional characteristics of bacteria, fungi, viruses, and archaea characteristics in children with obesity.

#### Box 1

To identify gut microbiota signatures that are characteristics of childhood obesity, we conducted a systematic review. We included all retrieved studies that reported on the gut microbiota of children with obesity, which included cross-sectional and case-control studies. In total, 63 studies (all were observational studies) that assessed changes in the gut microbiome of children with obesity were included in this review for analysis: 67% of studies ( $n = 42$ ) used 16S rRNA sequencing for microbial analysis,<sup>27–68</sup> nine studies used Real-time PCR (RT-PCR),<sup>69–77</sup> six studies used shotgun metagenomic sequencing,<sup>78–83</sup> and the remaining six studies used other methods to assess microbial differences.<sup>84–89</sup> Information on each study in this review is included in **Table S1** in the supplementary materials.

**Box 1 Figure.** Flow Diagram of Article Search and Selection Process.

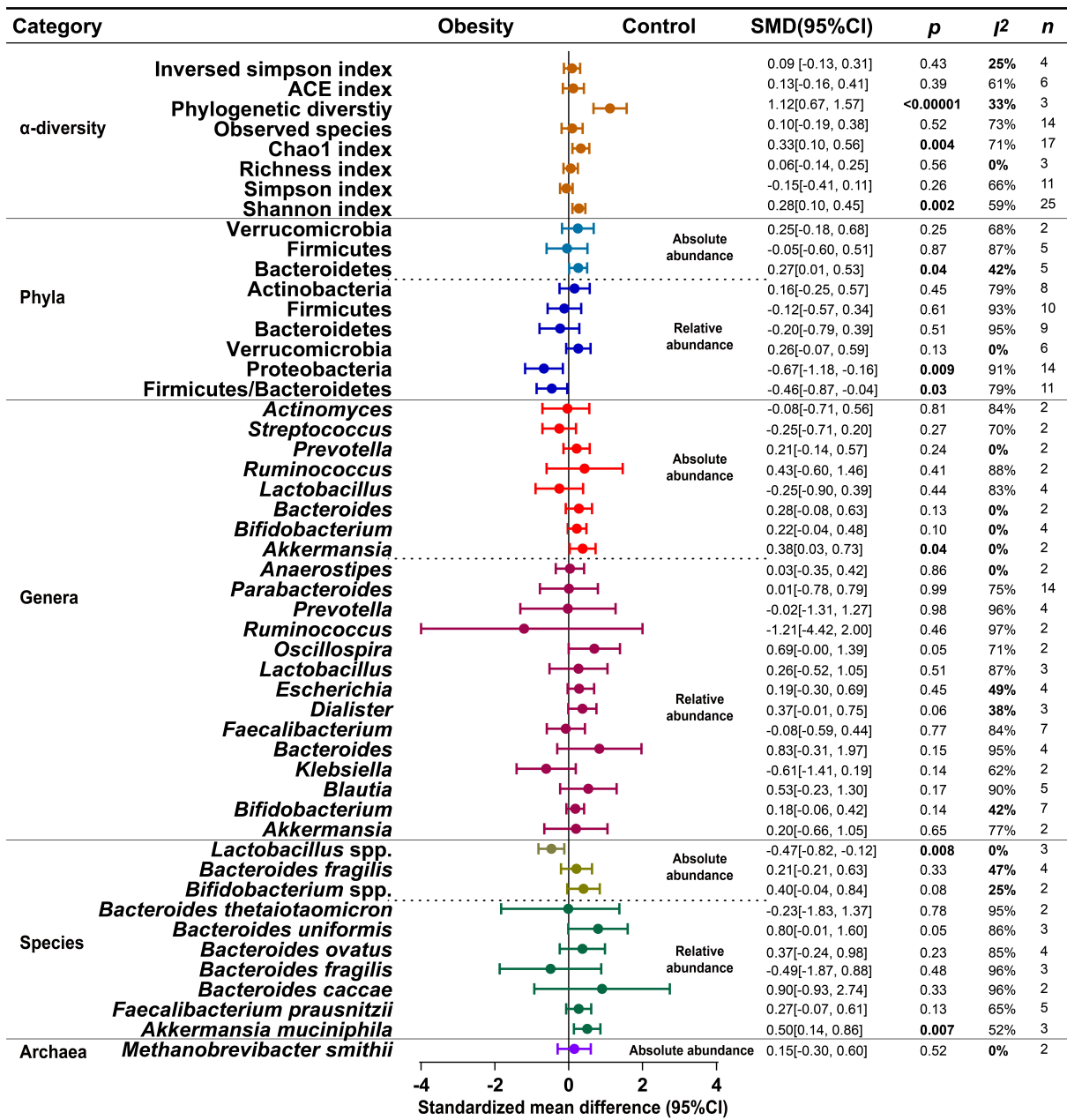


The literature search was conducted in July 2022, and an updated literature search was conducted in June 2024.

Summarized in Figure S1 are the main findings of the included studies regarding the relationship between microorganisms and childhood obesity. Only microbial features reported to correlate with obesity in three or more of the publications reviewed were included. The figure summarizes the percentage of included studies reporting the gut microbiota  $\alpha$ -diversity, bacteria (7 phyla, 30 genera, 12 species), and archaea

### 2.1. Microbial diversity

Of the 63 studies included, 65% (41/63) reported  $\alpha$ -diversity in children with and without obesity (Table S1). Eight indices were used to assess  $\alpha$ -diversity, and the most widely applied were Chao1, Shannon, Simpson and Observed species (Figure S1). There was no evidence of publication bias in any of the analyses (Figure S2). We conducted a meta-analysis based on the available



**Figure 1.** Meta-analysis of gut microbiota differences and α-diversity between obese children and controls. SMD, standardized mean difference. CI, confidence interval. The “p” indicates the level of significance. The “I<sup>2</sup>” indicates the results of the heterogeneity test. The “n” indicates the number of studies that provided usable data.

α-diversity index data (Figure 1 and Figure S3). The results revealed a statistically significant difference in Phylogenetic diversity, Chao1 index, and Shannon index of gut microbiota α-diversity index between children with obesity and control individuals. Compared to control, Phylogenetic diversity (6 studies with 3 estimates provided data, SMD = 1.12, 95% CI 0.67, 1.57, I<sup>2</sup> = 33%, p < 0.00001), Shannon index (32 studies with 25 estimates provided data, SMD =

0.28, 95% CI 0.10, 0.45, I<sup>2</sup> = 59%, p = 0.002), Chao1 index (22 studies with 17 estimates provided data, SMD = 0.33, 95% CI 0.10, 0.56, I<sup>2</sup> = 71%, p = 0.004) and were significantly decreased in children with obesity. Considering the heterogeneity, sensitivity analyses were performed by omitting each study in turn, which showed stability for the Chao1 index, and a decrease in heterogeneity for the Shannon index by omitting Méndez-Salazar 2018<sup>43</sup> (p = 0.002, I<sup>2</sup> = 42%)

(Figure S4). Based on the results of regional subgroup analyses, there were statistically significant differences for the Shannon index in Asian ( $p = 0.0002$ ,  $I^2 = 41\%$ ) and European ( $p = 0.01$ ,  $I^2 = 0\%$ ), and Simpson index ( $p = 0.001$ ,  $I^2 = 15\%$ ) in North America, with low heterogeneity. No significant differences or high heterogeneity were observed in any of the results based on age and BMI subgroup analyses (Figure S5).

Strong microbiota diversity is critical to the ability of the gut microbiota to adapt to stress and is a key indicator of good health. Meta-analysis showed a significant decrease in the Shannon index, Chao1 index and Phylogenetic diversity in obese children, aligning with the prevailing hypothesis. It is important to emphasize that different  $\alpha$ -diversity indices exhibit different microbiota characteristics. For example, while the Chao 1 index is based on the total number of bacteria within a community, the Shannon index and the phylogenetic diversity index also consider bacterial evenness and phylogenetic abundance, respectively. Therefore, despite heterogeneity between studies, including differences in region, gender and age, statistically significant differences between alpha diversity indices suggest that ecological disorder in obese children is expressed through disruption of phylogenetic abundance, bacterial homogeneity, and changes in bacterial numbers. However, the common assumption that “more diversity is better” may oversimplify the intricate complex mechanisms.<sup>90</sup> Measurement of diversity in children with obesity should be used as a starting point for further research of gut microecological mechanisms in obesity and will help to explore causal relationships between the gut microbiome and its host. Furthermore, the variation between the diversity of gut microbiota in obese children and adults deserves further exploration.<sup>91,92</sup>

Beta diversity comparison between childhood obesity and controls was reported in 43% (27/63) studies (Table S1). 17 studies reported consistent significant differences of obese versus non-obese children, while 10 studies reported conflicting results. The results of these studies suggest differences in the common phylogenetic structure of obese children compared to controls. However, the method of measurement, geographic region, age, and degree of obesity may influence the outcomes.

## 2.2. Variations in bacterial composition

Meta-analysis based on relative or absolute abundance data at the phyla level between childhood obesity and controls revealed statistically significant differences for Bacteroidetes, Proteobacteria and F/B ratios (Figure 1 and Figure S6). Compared to control individuals, F/B ratios (23 studies with 11 estimates provided data, SMD =  $-0.46$ , 95% CI  $-0.87$ ,  $-0.04$ ,  $I^2 = 79\%$ ,  $p = 0.03$ ) and relative abundance of Proteobacteria (21 studies with 14 estimates provided data, SMD =  $-0.67$ , 95% CI  $-1.18$ ,  $-0.16$ ,  $I^2 = 91\%$ ,  $p = 0.009$ ) were significantly increased, and absolute abundance of Bacteroidetes (36 studies with 5 estimates provided data, SMD =  $0.27$ , 95% CI  $0.01$ ,  $0.53$ ,  $I^2 = 42\%$ ,  $p = 0.04$ ) was significantly decreased in obese children. Considering the high heterogeneity, sensitivity analyses were performed by omitting each study in turn, with stable results (Figure S7). No publication bias in F/B ratios, relative abundance of Firmicutes and Proteobacteria (Figure S8). Based on the results of region, age and BMI subgroup analyses, there was a significant difference in F/B ratios ( $p < 0.00001$ ,  $I^2 = 0\%$ ) in European, and no significant difference between relative abundance of Firmicutes ( $p = 0.89$ ,  $I^2 = 0\%$ ) in school-aged children and absolute abundance of Firmicutes ( $p = 0.63$ ,  $I^2 = 0\%$ ) in overweight children, with high heterogeneity of the results of the analyses of the other subgroups (Figure S9).

At the genera level, the absolute abundance of *Akkermansia* (6 studies providing 2 estimates, SMD =  $0.38$ , 95% CI  $0.03$ ,  $0.73$ ,  $I^2 = 0\%$ ,  $p = 0.04$ ) was significantly decreased in obese children only compared with controls (Figure 1 and Figure S10). Sensitivity analysis showed a significant increase relative abundance of *Faecalibacterium* by omitting Tian 2023<sup>65</sup> ( $p = 0.0004$ ,  $I^2 = 0\%$ ) (Figure S11). Region subgroup analyses showed low heterogeneity and significant increase relative abundance of *Faecalibacterium* in North America ( $p = 0.02$ ,  $I^2 = 21\%$ ) (Figure S12).

At the species level, *Akkermansia muciniphila* (4 studies) was consistently decreased in children with obesity (Figure S1). Meta-analysis results showed that relative abundance of *Akkermansia muciniphila* (4 studies providing 3 estimates, SMD =  $0.50$ , 95% CI  $0.14$ ,  $0.86$ ,  $I^2 = 52\%$ ,  $p = 0.007$ ) was

significantly decreased and absolute abundance of *Lactobacillus* spp. (3 studies providing 3 estimates, SMD = -0.47, 95% CI -0.82, -0.12,  $I^2 = 0\%$ ,  $p = 0.008$ ) was significantly increased in obese children compared to controls (Figure 1 and Figure S13). Sensitivity analyses showed a significant decrease relative abundance of *A. muciniphila* ( $p < 0.0001$ ,  $I^2 = 0\%$ ) and *Bacteroides ovatus* ( $p < 0.0001$ ,  $I^2 = 0\%$ ) after omitting López-Contreras 2018<sup>45</sup> (Figure S14). Region subgroup analyses demonstrated low heterogeneity and a significant decrease in the relative abundance of *Faecalibacterium prausnitzii* among Asian individuals ( $p = 0.0002$ ,  $I^2 = 21\%$ ) (Figure S15).

Clinical studies have demonstrated notable benefits in overweight/obese individuals supplementing with pasteurizing *A. muciniphila* bacteria to significantly improve insulin resistance, blood cholesterol and inflammatory factor levels.<sup>93</sup> It is estimated that *F. prausnitzii* makes up 1% to 6% of the total microbiota of healthy individuals,<sup>94</sup> and has been shown to impart anti-inflammatory characteristics in part by the production of butyrate.<sup>94,95</sup> However, these anti-inflammatory benefits are not observed in children with obesity. Despite presumably sharing a core microbiome architecture, recent research has indicated that the gut microbiota of children between the ages of 3 and 18 is distinct in terms of both function and taxonomy compared to that of adults.<sup>20</sup> This suggests that the establishment of the gut microbiota may require a somewhat lengthy period. A meta-analysis of data from adults with obesity revealed that the abundance of *Dorea*, *Dialister*, *Fusobacterium*, *Sutterella*, *Streptococcus*, and *Prevotella* was significantly increased in obesity. Conversely, *Bifidobacterium* exhibited a lower abundance.<sup>92</sup> In contrast, in children with obesity, *Dialister*, *Streptococcus*, and *Prevotella* did not observe a similar pattern of changes as in adults with obesity (Figure 1), indicating a disparity between the gut microbiota of children and adults with obesity.

### 2.3. Variations in fungi composition

Human gut fungi are receiving increased research attention<sup>96,97</sup> due to their potential involvement in the etiology of numerous gut-associated diseases.<sup>98</sup>

*Candida* is a diploid fungus that causes disease by invading the host's mucous membranes and tissues, triggering local and systemic inflammatory responses.<sup>99</sup> There are many species of fungi in the genera *Candida*, but only a few are pathogenic to humans, mainly *Candida albicans*, *Candida tropicalis*, *Candida parapsilosis*, and *Candida dubliniensis*.<sup>100</sup> Only 5% (3/63) of the systematically reviewed studies investigated the differences in fungi composition between children with and without obesity. As there were fewer than three studies for the same fungi, these studies were not included in Figure S1. The study by Grigorova *et al.*<sup>51</sup> reported that *Candida* was not detected in any of the tested feces specimens from children with obesity, while in the control group three *Candida* species were detected: *C. albicans*, *Candida glabrata*, and *Candida krusei*. Borgo *et al.*<sup>88</sup> also found a relatively low abundance of *C. albicans* and *Saccharomyces cerevisiae* in children with obesity compared to the normoweight children. Furthermore, *C. parapsilosis* and *C. glabrata* were detected in normoweight children but not children with obesity. These findings are contrary to research in mice that show *C. parapsilosis* stimulates the formation of fatty acids by producing a fungal lipase that causes obesity.<sup>101</sup> Peng *et al.*<sup>38</sup> report that *Basidiomycota* sp. was more abundant in children with obesity. Reporting gut mycobiota in human fecal specimens and its presence and relative abundance is currently nascent without a consensus on what is considered an "ideal mycobiome".<sup>102</sup>

Previously, in vitro culturing was the major method used by researchers to study fungi from the intestinal environment. Despite recent breakthroughs in molecular approaches that eliminate the need to cultivate bacteria, this research has mostly concentrated on the non-fungi component of this ecosystem. Consequently, few studies have included the gut mycobiome and even fewer in the pediatric population. It is noteworthy to emphasize that the gut is a diverse and intricate ecosystem within the human body. Research findings indicate that the most prevalent genera in the mycobiome of a healthy adult gut include *Candida*, *Saccharomyces*, and *Cladosporium*.<sup>103</sup> *Candida* spp. continues to be identified as the prevailing fungi species among individuals aged 65 years and older.<sup>104</sup> Nevertheless, in diseases such as

obesity<sup>105</sup> and inflammatory bowel disease,<sup>106</sup> an increase in *Candida* spp. and its inverse correlation with bacterial microbiome diversity have been repeatedly observed. Some studies suggest that *Candida* spp. may break down starch in foods, releasing simple sugars that are fermented by other bacteria.<sup>107</sup> Furthermore, *C. albicans* was observed to decrease the levels of dissolved oxygen in its immediate environment, creating favorable conditions for the proliferation of anaerobic bacteria, such as *Clostridioides difficile*.<sup>108,109</sup> Overall, observational and exploratory investigations of gut fungi in children with obesity have highlighted the significant role of dysbiosis of the gut microbes in the pathogenesis of obesity. Understanding cross-kingdom microbial interactions between gut microorganisms in pediatric obesity is important for elucidating the relationship between obesity and microbiota.

#### 2.4. Variations in viruses composition

Human feces contain approximately  $10^8$  to  $10^9$  viruses per gram of feces with DNA bacteriophages predominating and RNA viruses being only a minor fraction of the gut.<sup>110,111</sup> Maya-Lucas *et al.*<sup>78</sup> noted that the relative abundance of human herpesvirus 4 (NC-007605.1) increased from 0.08% in normoweight children to 1.53% in children with obesity, while Torque teno midi virus 1 (NC-009225.1) decreased from 2.21% in normoweight children to 0.72% in children with obesity. Furthermore, specific virus particles, notably bacteriophages, might play an essential role in the maintenance of certain strains of gut bacteria associated with the development of obesity.<sup>112</sup> Shirley *et al.*<sup>113</sup> used metagenomic sequencing to evaluate virus-like particles (VLPs) in the feces of children with obesity. The results indicate an increase in the prevalence and variety of all phage categories in children with obesity and metabolic syndrome. In the same cohort of children, it was observed that the virome of children with obesity was predominantly dominated by Caudovirales, with only Inoviridae showing a significant increase in the number of viruses.<sup>114</sup> Intriguingly, further analysis of the enteroviruses of

the same cohort of children revealed that loss of CrAssphage stability was associated with obesity, with the effect being more pronounced in cases of obesity accompanied by metabolic syndrome.<sup>115</sup> However, additional research is required to comprehend the underlying reasons behind the decrease in CrAssphage variety and abundance in individuals with obesity and metabolic syndrome, which contradicts the observed patterns in all phagesomes.

The enteric viruses and phage-bacteria interaction are also crucial for health effects. According to a study by Dutilh *et al.*,<sup>116</sup> “CrAss-like” phages can infect Bacteroidetes taxa such as *Prevotella intermedia* and *Bacteroides* spp. and persist inside these hosts. Phages can coexist with gut bacteria for long periods and have the potential to modulate the gut microbiota.<sup>117</sup> Furthermore, it has been suggested that successful treatments against *Clostridioides difficile* utilizing bacteria-free fecal filtrate imply that phagosome modification may be a useful therapeutic technique to stabilize bacterial eubiotics in the microbiome.<sup>118</sup> As in other niches, phages play a crucial part in childhood obesity and the disorders that accompany it. Although the nature of all viruses and their role in the health or dysbiosis states of the intestinal microbiome are not currently known, the evidence indicates that their role may be fundamental for bacterial dynamics, pointing to the need to perform deep gut virome studies to elucidate their ecological role in this niche.

#### 2.5. Variations in archaea composition

Methanobacteriales are the most abundant order of archaea and their members can produce methane, reduced CO<sub>2</sub> or methanol with H<sub>2</sub> as the main electron donor. The main methanogens in the gut microbiota are *Methanobrevibacter smithii*,<sup>119</sup> *Methanosphaera stadtmanae*,<sup>120</sup> and *Methanomassilicoccus luminyesis*.<sup>121</sup> Animal studies have suggested that archaea, specifically *M. smithii*, may play a role in the development of obesity. Samuel and Gordon<sup>122</sup> observed that *M. smithii* played a critical role in facilitating an increased capacity of *B. thetaiotaomicron* to

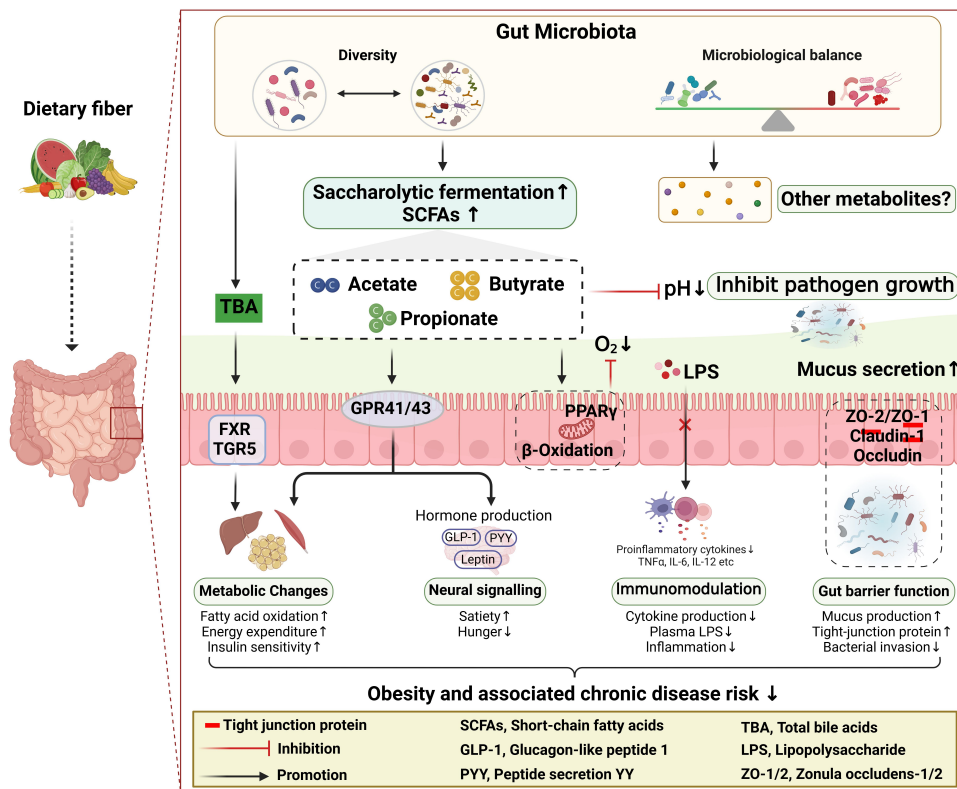


digest polyfructose-containing glycans leading to increased production of SCFAs and total liver triglycerides in mice.

However, meta-analysis showed no statistically significant difference in the absolute abundance of *Methanobrevibacter smithii* (3 studies providing 2 estimates, SMD = 0.15, 95% CI -0.30, 0.60,  $I^2 = 0%$ ,  $p = 0.52$ ) between obese children and controls (Figure 1). In addition, there was an increase in the relative abundance of a different unclassified species of *Methanobrevibacter* spp. in children with obesity.<sup>78</sup> Members of this genera have productive saccharolytic activity, allowing polysaccharide digestion in the gut.<sup>123</sup> Although the reviewed studies have not yielded similar findings as observed in animals, it should be noted that this does not negate the potential significance of methanogens in metabolic dysregulation among children with obesity. However, it is important to acknowledge that the current data available are limited and further research is warranted to elucidate their precise role in obesity.

## 2.6. Childhood obesity and gut microbiota: outstanding questions

In this review report, although we identified some gut microbiota characteristics of pediatric obesity, it is important to note that our target population is in broad and diversified in terms of physiological developmental stages. The reported results exhibit a high degree of heterogeneity, with confounding factors such as diet, geography, age, sample size, and testing methods often influencing the composition of gut microbiota. Furthermore, whether changes in the gut microbiota precede the development of obesity in children or if they reflect the obese phenotype as a whole remains to be answered. The interrelationship between nutrition, microbiota, genetic factors, immunity, host life behaviors, and the development of childhood obesity is complex, making it challenging to provide definitive conclusions on this subject. Furthermore, the primary succession of the gut microbiota in the human life cycle ends with the formation of the climacteric community, which is



**Figure 2.** A summary of the mechanisms by which DFs initiate metabolic changes to combat the development of childhood obesity. These mechanisms include the production of microbial metabolic products, noting SCFAs, a decrease in luminal pH and O<sub>2</sub>, regulation of the immune system (modulating cytokine production), improving gut barrier function, promoting satiety through gut-brain signaling, and enhancing oxidative metabolism.

thought to be achieved during adolescence and largely maintained into adulthood; this community is characterized by relative stability.<sup>18,124</sup> Therefore, this remains a very challenging area of research, not only to determine the causal relationship between gut microbiota and obesity but also to propose potential effective interventions that can host a healthier microbiota from childhood and thus promote a better metabolic balance of the organism.

### 3. Effects of dietary fibers on the gut microbiota of children with obesity

#### 3.1. Microbial metabolism of dietary fibers and obesity implications

DFs serve as a crucial energy source for microorganisms residing in the gastrointestinal tract. Under specific intestinal conditions, anaerobic bacteria ferment complex carbohydrates by activating mechanisms consisting of critical enzymes and metabolic pathways that produce metabolites such as SCFAs (Figure 2).

SCFAs are organic compounds composed predominantly of acetate, propionate, and butyrate, and one of their mechanisms of action in obesity is to regulate intestinal endocrine signals, influencing glucose and lipid metabolism. Acetate, butyrate, and propionate can interact with G protein-coupled receptors (GPCR41, GPCR43)<sup>125–127</sup> leading to the activation of anorexigenic hormones such as glucagon-like peptide 1 (GLP-1) and peptide secretion YY (PYY).<sup>128,129</sup> These hormones play a crucial role in regulating appetite, reducing energy intake and helping to promote glucose metabolism.<sup>130</sup> SCFAs further improve oxidative metabolism and insulin sensitivity in the liver and adipose tissue by interacting with the G protein-coupled receptor protein FFAR3 and promoting uncoupling protein 2 (UCP2) activity.<sup>131</sup> Another mechanism of action of SCFA in improving obesity is the reduction of metabolic endotoxemia and inflammation. Obesity is thought to lead to a “leaky gut” and subsequent metabolic endotoxemia (increased serum Lipopolysaccharides [LPS] levels) in animal models and to some extent in humans.<sup>132</sup> LPS appears to bridge the gap between the gut microbiota and

inflammation, and once it is translocated into systemic circulation, it binds to Toll-like receptor 4 and initiates the production of pro-inflammatory cytokines.<sup>133</sup> SCFAs also play an anti-inflammatory role by regulating the size and function of the colonic regulatory T cells, leading to a decrease in pro-inflammatory factors (e.g., IL-6, IL-8) in intestinal epithelial cells and metabolic tissues (e.g., adipose tissue).<sup>134</sup> Furthermore, SCFAs play a crucial role in preserving the integrity of the intestinal mucus barrier and creating a favorable milieu that supports the proliferation of putatively beneficial gut microbes. *Bacteroides thetaiotaomicron* produces acetate and propionate, which promote differentiation of goblet cells and stimulate mucin secretion. In contrast, *F. prausnitzii* utilizes acetate to generate butyrate, which attenuates the effect of *B. thetaiotaomicron* on mucus and helps the epithelial cells to maintain the proper ratio of different types of cells in the secretory lineage, thereby reducing mucus overproduction and protecting the intestinal mucus barrier.<sup>135</sup> Butyrate stimulates  $\beta$ -oxidation in mitochondria by activating the nuclear receptor peroxisome proliferator-activated receptor (PPAR) in colonocytes, which maintains anaerobic conditions in the intestinal lumen.<sup>136</sup> SCFAs also decrease the pH of the intestinal lumen and prevent the growth of pathobionts.

Collectively, these acids serve as an additional energy source and concurrently regulate various systemic processes by activating signaling cascades within the host. However, it is important to note that observational studies conducted on human obesity have often have frequently yielded disparate findings regarding the quantification of SCFAs in feces or blood. For instance, one study indicated that obese children and adolescents harbor a higher abundance of carbohydrate-fermenting bacteria in their microbiota compared to normoweight counterparts, thereby augmenting SCFAs biosynthesis rates and providing the host with an additional energy source, ultimately stored as lipids or glucose.<sup>74</sup> This apparent paradoxical observation implies that in addition to the SCFAs levels made by some members of the gut microbiota, there might be a direct recognition of specific bacterial components by some GPCRs in the host, which in turn modulate other metabolic processes that may

contribute to obesity. Furthermore, it is important to note that the physiological mechanisms of SCFAs have primarily been studied in animal models and require further validation through clinical trials.

DFs may also further influence clinical outcomes in obesity by modulating the gut microbiota, thereby affecting the metabolism of the bile acid pool in the gut. Bile acids (BAs) are cholesterol metabolites produced in hepatocytes and specific gut microbes, such as *Lactobacillus*, *Bifidobacterium*, and *Bacteroides*, that can metabolize into secondary BAs through processes of depolymerization, dehydroxylation, and repolymerization.<sup>137</sup> Disturbances in the gut microbial population prevent the conversion of primary bile acids, leading to their accumulation and a decrease in secondary bile acids. Primary bile acids act more specifically on G protein-coupled receptors (TGR5), whereas secondary bile acids primarily act on the Farnesoid X receptor (FXR).<sup>138</sup> The activation of these receptors has been shown in animal models to cause changes in lipid and carbohydrate metabolism, energy expenditure, and inflammation.<sup>137</sup>

### 3.2. Role of dietary fibers in the management of the gut microbiome in children with obesity

As mentioned above, there is evidence that children with obesity have unique gut microbiota signatures that differ from that of healthy children.<sup>21,139</sup> Therefore, whether nutritional interventions can redress microbial imbalances in pediatric obesity has garnered increasing attention in the field of nutrition. It is commonly recognized that a healthy diet is a critical environmental factor mediating the composition and metabolic function of the gut microbiota. DFs modulate the gut microbiome primarily including three strategies: 1) with more complex dietary patterns that provide rich sources of intrinsic DFs; 2) with a symbiotic mixture of live microorganisms and selective prebiotics fiber(s); 3) with the isolated fiber(s). The impact of diet on the composition of the gut microbiota is summarized in Table 1.

#### 3.2.1. Effects of intrinsic dietary fibers and high-fibers diet pattern on the gut microbiota of children with obesity

As depicted in Figure 1, Actinobacteria, *Prevotella*, *Bifidobacterium*, *Dialister*, *Escherichia*, *Anaerostipes*, *Akkermansia*, *Bacteroides* and  $\alpha$ -diversity metrics other than the Simpson index showed a tendency to be less prevalent in children with obesity, and although there was no significance, inclusion in the Meta-analysis reported low heterogeneity. In high DFs dietary patterns 11% (1/9) of studies reported an increase in the Shannon index, ACE index and Chao1 index and 11% of studies (1/9) reported a decrease in Chao1 index, Observed OTUs, Phylogenetic diversity and Shannon index. An increase in *Prevotella* is associated with long-term dietary patterns, complex carbohydrates, or plant-based foods.<sup>150</sup> Many of the species that respond to changes in traditional diet intake (high plant-based and low-fat foods) seem to belong to Actinobacteria, which are nutritionally specialized.<sup>151,152</sup> Actinobacteria (such as *Bifidobacterium*, *Bifidobacterium* spp.) few genera increase in response to high-fiber diets.<sup>143,145–147</sup> In response to the Western diet, the abundance of *Bacteroides* and *Klebsiella* increased, which was accompanied by a decrease in *Prevotella*,<sup>141,142,153</sup>

Recently, there has been an increasing recognition that consuming DFs naturally present in food (in contrast to consuming isolated DFs as supplements) may be most effective at regulating gut bacteria and enhancing overall health.<sup>154</sup> Regarding dietary interventions for childhood obesity, it is important to consider not only whether the gut microbiota is being modulated, but also the improvement of metabolism in children. SCFAs, as key metabolites of fermentation of DFs by the gut microbiota, have a broad impact on various aspects of host physiology,<sup>155</sup> Table 1 shows that only one cross-sectional study investigating high-fiber diets provided evidence of increased SCFAs.<sup>144</sup> A traditional grain diet improved weight, BMI, and liver function metabolism in children with obesity, according to another study.<sup>146</sup> It is important to note that while the effects of high-fiber diets on the body's metabolism should be examined in relation to the microbiota, these diets may also affect the metabolism directly, independent of the

Table 1. Studies in which the effects of different DFs on gut microbiota in children with obesity was assessed.

|  |                            | Study   |                   | Methodology |                      | Microbiota             |            | Microbial Diversity   |                                      | Metabolic Effects   |                                    | References |  |
|--|----------------------------|---|-------------------|-------------|----------------------|------------------------|------------|---|--------------------------------------|---|------------------------------------|------------|--|
| Trial design   | Population                 | Dose  | Age               | n           | Duration             | Methodology            | Microbiota | Microbiota  | Diversity                            | Metabolic Effects   | References                         |            |  |
| <b>Intrinsic DFs and high-fiber diet pattern</b>   |                            |   |                   |             |                      |                        |            |   |                                      |   |                                    |            |  |
| Low fruit and rich in meat   | Cross-sectional study      | Patients with Prader-Willi syndrome (overweight or obesity) | 23.1 ± 13.4 g/day | 5-8         | 31                   | 4-day food diary       | 16S rRNA   | <i>Klebsiella</i> ↑<br><i>Eubacterium</i> ↑<br><i>Lactobacillus</i> ↑<br><i>Lachnospiridium</i> ↓<br><i>Murimonas</i> ↓<br><i>Alistipes</i> ↓<br><i>Prevotella</i> ↓<br><i>Bacteroides</i> ↑<br><i>Prevotella</i> ↑<br><i>Ruminococcus</i> ↑<br><i>Blautia</i> ↑<br><i>Bacteroides</i> ↑<br><i>Roseburia</i> ↑<br><i>Lachnospira</i> ↑<br><i>Slackia</i> ↓<br><i>Prevotella</i> ↓<br><i>Oscillospira</i> ↓<br><i>Faecalibacterium</i> ↑<br><i>Eubacterium</i> ↑<br><i>Roseburia</i> ↑<br><i>Lachnospiraceae</i> ↑<br><i>Peptostreptococcaceae</i> ↑ | Shannon index↑                       | <i>Klebsiella</i> are significantly positively correlated with total and LDL-cholesterol, <i>Alistipes</i> are significantly negatively correlated with body fat mass, total and LDL-cholesterol levels, fasting insulin levels and HOMA-IR. Fructose intake is negatively associated with the genera <i>E. eligens</i> and <i>Streptococcus thermophilus</i> | Garcia-Ribera et al <sup>140</sup> |            |  |
| 51.6% total carbohydrates, 21.9% total sugar, 5.1% fiber, 31.8% total fat, 16.5% total protein | Cross-sectional study      | Overweight or Obesity                                       | 16.1 ± 6.7 g/day  | 12-19       | 52                   | 24-hour diet recalls   | 16S rRNA   | <i>Bacteroides</i> ↑<br><i>Blautia</i> ↑<br><i>Roseburia</i> ↑<br><i>Lachnospira</i> ↑<br><i>Slackia</i> ↓<br><i>Prevotella</i> ↓<br><i>Oscillospira</i> ↓<br><i>Faecalibacterium</i> ↑<br><i>Eubacterium</i> ↑<br><i>Roseburia</i> ↑<br><i>Lachnospiraceae</i> ↑<br><i>Peptostreptococcaceae</i> ↑   | none stated                          | Fructose intake is negatively associated with the genera <i>E. eligens</i> and <i>Streptococcus thermophilus</i>  | Jones et al <sup>141</sup>         |            |  |
| High carbohydrate/high fat diet or high protein/high fat diet                                  | Prospective study          | Overweight or Obesity                                       | None stated       | 2-9         | 707                  | 7-day diet recalls     | 16S rRNA   | <i>Bacteroides</i> ↑<br><i>Roseburia</i> ↑<br><i>Lachnospira</i> ↑<br><i>Slackia</i> ↓<br><i>Prevotella</i> ↓<br><i>Oscillospira</i> ↓<br><i>Faecalibacterium</i> ↑<br><i>Eubacterium</i> ↑<br><i>Roseburia</i> ↑<br><i>Lachnospiraceae</i> ↑<br><i>Peptostreptococcaceae</i> ↑   | none stated                          | Inflammatory (IL-15, TNF-α, IP-10, IL6, IL-8) ↑   | Rampelli et al <sup>142</sup>      |            |  |
| Higher fiber and nonstarch polysaccharides diet  | Cross-sectional study      | Overweight or Obesity                                       | 18 g/day          | 5           | 319                  | one month food diary   | 16S rRNA   | <i>Faecalibacterium</i> ↑<br><i>Eubacterium</i> ↑<br><i>Roseburia</i> ↑<br><i>Lachnospiraceae</i> ↑<br><i>Peptostreptococcaceae</i> ↑   | Shannon index↑                       | None stated   | Leong et al <sup>143</sup>         |            |  |
| High traditional vegetable-based, low fat, sugar diets   | Cross-sectional study      | 28 rural children (Buriram)                                 | None stated       | 9-11        | 45                   | 7-day dietary records  | 16S rRNA   | <i>Faecalibacterium</i> ↑<br><i>Bifidobacterium</i> ↑<br><i>Clostridium</i> ↑<br><i>Collinsella</i> ↑<br><i>Bifidobacterium catenulatum</i> ↑<br><i>B. longum</i> ↑<br><i>Bacteroides</i> ↓<br><i>Ruminococcus</i> ↓<br><i>Blautia</i> ↓<br><i>Escherichia</i> ↓<br><i>Bifidobacterium spp.</i> ↑<br><i>B. pseudocatenulatum</i> ↑  | ACE index↑<br>Chao1 index↑           | Parabacteroides ↑<br>Butyrate ↑   | Kisuse et al <sup>144</sup>        |            |  |
| Whole grains, traditional Chinese medicinal foods, and prebiotics diet                         | Dietary intervention trial | Prader-Willi syndrome and obesity                           | 50 g/day          | 3-16        | 360, 30, 60, 90 days | Metagenomic sequencing |            | <i>Faecalibacterium</i> ↑<br><i>Bifidobacterium</i> ↑<br><i>Clostridium</i> ↑<br><i>Collinsella</i> ↑<br><i>Bifidobacterium catenulatum</i> ↑<br><i>B. longum</i> ↑<br><i>Bacteroides</i> ↓<br><i>Ruminococcus</i> ↓<br><i>Blautia</i> ↓<br><i>Escherichia</i> ↓<br><i>Bifidobacterium spp.</i> ↑<br><i>B. pseudocatenulatum</i> ↑  | None stated                          | None stated   | Li et al <sup>145</sup>            |            |  |
|  |                            |   |                   | 38          |                      |                        |            | Chao1 index↓<br>Observed OTU↓<br>Phylogenetic diversity↓<br>Shannon index↓<br>gene richness↓  | Body weight↓<br>BMI↓<br>ALT↓<br>AST↓ | Zhang et al <sup>146</sup>  |                                    |            |  |

(Continued)

Table 1. (Continued).

|  | Study   | Population            | Dose        | Age  | n            | Duration | Methodology                | Microbiota   | Microbial Diversity  | Metabolic Effects   | References                          |
|--|---|-----------------------|-------------|------|--------------|----------|----------------------------|--|--|---|-------------------------------------|
| Proteins and complex carbohydrate patterns                             | Cross-sectional study   | Overweight or Obesity | None stated | 6-12 | 46           | \        | Metagenomic sequencing     | <i>Holdemania spp.</i> ↑<br><i>Coprococcus catus</i> ↓   | none stated  | z-BMI ↓<br>waist circumference ↓<br>hip circumference ↓                                     | Orbe-Orihuela et al <sup>147</sup>  |
| Whole grains, traditional Chinese medicinal foods, and prebiotics diet | Dietary intervention trial  | Overweight or Obesity | 50 g/day    | 3-18 | 8730, 60, 90 | 165 days | rRNA                       | <i>Bifidobacterium</i> ↑<br><i>Lactobacillus</i> ↑   | none stated  | None stated   | Hou et al <sup>68</sup>             |
| <b>Synbiotic</b>   |   |                       |             |      |              |          |                            |  |  |   |                                     |
| Synbiotic (FOS)  | Randomized triple-masked controlled trial   | Obesity               | one capsule | 6-18 | 70           | 8 weeks  | surface-cultured of plates | <i>Lactobacillus</i> ↑<br><i>Bifidobacteria</i> ↑  | none stated  | Weight ↓<br>Body mass index ↓<br>Z-score ↓<br>IL-6 ↓<br>TNFα ↓<br>Hs-CRP ↔<br>Adiponectin ↑ | Kelishadi et al <sup>140</sup>      |
| Synbiotic (FOS)  | Single-center, prospective, randomized, double-blind, placebo-controlled clinical study | Obesity               | 1 sachet    | 8-17 | 54           | 12 weeks | 16S rRNA                   | Compared to baseline:<br>Bacteroidetes ↑<br><i>Prevotella</i> ↑<br><i>Dialister</i> ↑<br>F/B ratio ↓<br>Compared to placebo groups: <i>Collinsella stercoris</i> species were dominant in the symbiotic group  | Chao1 index ↔<br>Observed ASV ↔<br>Simpson index ↔<br>Shannon index ↔                  | None stated   | Kilic Yildirim et al <sup>148</sup> |
| <b>Isolated fiber</b>  |   |                       |             |      |              |          |                            |  |  |   |                                     |
| Inulin   | Double-Blind, RCT   | Obesity               | 8 g/day     | 7-12 | 42           | 16 weeks | 16S rRNA qPCR              | <i>Actinobacteria</i> ↑<br><i>Bifidobacterium</i> ↑<br><i>Collinsella</i> ↑<br><i>B. adolescentis</i> ↑<br><i>B. longum</i> ↑<br><i>Ruminococcus</i> ↓<br><i>F. prausnitzii</i> ↓<br><i>E. eligens</i> ↓<br><i>B. vulgatus</i> ↓<br><i>R. gaurvrauii</i> ↓ | Shannon index ↑<br>Simpson index ↔<br>observed OTUs ↑<br>beta-diversity ( $p > 0.05$ ) | Body Weight ↓<br>Body Fat (%) ↓<br>trunk fat ↓<br>IL-6 ↓<br>TAG ↓<br>primary bile acids ↔   | Nicolucci et al <sup>149</sup>      |

microbiota. Foods such as those replaced by DFs (if they replace monosaccharides) produce additional benefits.

It is tempting to target ecological markers similar to those found in children with healthy diets (higher richness and diversity, stimulation of beneficial microbes), but it is unclear which microbiota response in children should be supported. In conclusion, most studies have shown a strong association between dietary patterns, gut microbiome, and childhood obesity.

### **3.2.2. Effects of synbiotics mixture (live microorganisms and selective prebiotics fibers) on the gut microbiota of children with obesity**

Synbiotic mixtures are a combination of prebiotics and probiotics that may be used to moderate dysfunctional gut microbiomes.<sup>156</sup> The effect of synbiotics on  $\alpha$ -diversity was not observed in the included studies (only 1 of the 2 studies reported relevant results). Kelishadi<sup>140</sup> and colleagues used synbiotics capsules (fructooligosaccharides [FOS] plus *Lactobacillus* and *Bifidobacteria*) to intervene in children with obesity for eight weeks and found significant increases in *Lactobacillus*, as well as significant reductions BMI, IL-6, and tumor necrosis factor- $\alpha$  (TNF $\alpha$ ). Another study used a Synbiotics formulation (probiotics mixture including *Lactobacillus acidophilus*, *Lactocaseibacillus rhamnosus*, *Bifidobacterium bifidum*, *Bifidobacterium longum*, *Enterococcus faecium* (total  $2.5 \times 10^9$  CFU/sachet) and FOS (625 mg/sachet) in children with obesity over a 12-week period; compared to baseline, Bacteroidetes, *Prevotella*, and *Dialister* significantly increased, F/B ratio and BMI significantly decreased.<sup>148</sup> Interestingly, Bacteroidetes was significantly decreased and *Dialister*, *Lactobacillus* showed a trend of reduction in children with obesity. These data suggest that synbiotics can drive changes in the gut microbiota of obese children, with the potential to increase the diversity of the gut microbiota. However, synbiotics formulations often require higher than expected dosages of prebiotics for practical use to ensure that substrates are available for both the resident microbiota and the co-administered microbes in a highly competitive microenvironment

within the gut.<sup>157–159</sup> Therefore, Precise understanding of the synergistic impact of probiotics and prebiotics in synbiotics, as well as the mechanism by which they regulate the gut microbiota in children with obesity, remains challenging within the realm of precision nutrition.

### **3.2.3. Effects of isolated dietary fibers on the gut microbiota of children with obesity**

Isolated DFs can have a significant impact on the composition, diversity and abundance of the microbiome, providing a wealth of substrates for fermentation reactions carried out by specific microbial species.<sup>160</sup> Microorganisms possess the necessary enzymes to hydrolyze a wide variety of complex carbohydrates, including glycoside hydrolases, glycosyltransferases, polysaccharide lyases, and carbohydrate esterases.<sup>161</sup> As a result, having a variety of DFs (e.g., cellulose, inulin, fructans, resistant starches) in a diet that contains a range of monosaccharide units and  $\alpha$  and  $\beta$  linkages is more supportive of a diverse gut microbial community than a diet with a less diverse substrate load (e.g., refined diets).<sup>162</sup> Despite the growing body of literature on the ability of isolated DFs to positively improve childhood obesity,<sup>163,164</sup> there is limited information on their impact on microbial diversity and abundance. Several studies have demonstrated that the intake of isolated DFs shapes the gut microbiota of children with obesity.<sup>149,165</sup> In children with overweight or obesity, 16 weeks of oligofructose-enriched inulin (8 g/day) increased  $\alpha$ -diversity (Shannon index, Observed OTUs), associated with an increase in Actinobacteria (such as *Bifidobacterium*) and a decrease in Firmicutes (such as *Ruminococcus*).<sup>149</sup> Meanwhile, at the level of OTUs, *F. prausnitzii* decreased significantly. These changes in microbiota outcomes suggest that DFs are better at improving gut microbiota in obese children (Figure 2). Moreover, this prebiotics intervention improved children's body weight, body fat, IL-6 levels, and triglyceride content. These beneficial effects of isolated DFs on childhood obesity aspects are thought to be at least partially mediated by the microbiota induced changes but more direct evidence still has to be provided.

In summary, intrinsic dietary, synbiotics, and isolated fibers exert specific modulatory effects on the gut microbiota composition and  $\alpha$ -diversity in children with obesity. Current evidence does not support that DFs can modulate the bacteria (e.g., *Akkermansia*, *A. muciniphila*) found to be significantly decreased in children with obesity in the systematic review. *A. muciniphila*, a mucin-degrading bacterium, has a negative association with obesity, diabetes, and colitis.<sup>166</sup> Supplementation with oligofructose has been shown to increase the number of goblet cells in animals, which subsequently exerts a prebiotic function on *A. muciniphila*.<sup>167</sup> High-fermentable (pectin) fiber also modulates the gut immune environment in mice, which in turn promotes *Akkermansia* enrichment,<sup>168</sup> and the findings suggest that the fiber effect on *Akkermansia* may be independent of fermentation but rather by improving the gut microecological environment. Furthermore, low-fermentable fibers augment crypt length, goblet cell maturation, and mucin secretion via mechanical stimulation caused by increased stool bulk.<sup>169</sup> Cellulose (an insoluble fiber with low-fermentable) supplementation significantly increased *Akkermansia* abundance in mice with colitis, potentially by promoting mucin secretion from goblet cells.<sup>170</sup> Low-fermented fibers effectively accelerate gut transit time, which may have implications for regulating the structure of the gut microbiota.<sup>171</sup> However, there is a lack of clinical evidence investigating the impact of DF supplementation on *Akkermansia* in children with obesity, particularly the effect of isolated low-fermentable fiber on gut microbiota in this population.

### **3.2.4. Dietary fibers and gut microbiota in children with obesity: remaining unsolved questions**

The identification and implementation of suitable nutritional intervention techniques for modulating the gut microbiota in children with obesity is a prominent subject of interest for forthcoming scientific investigations. For many years, the study of DF-microbiota interactions using a reductionist approach has elucidated DFs dependent and independent effects on specific microbiota.<sup>172</sup> Currently, dietary patterns with high DFs are an important strategy in nutritional

intervention programs. As shown in [Table 1](#), the more complex dietary interventions assessing the effect of DFs on the gut microbiota in children with obesity provide relatively more evidence, but substantial evidence is still lacking in general (such as randomized clinical trials). Further investigation is required to ascertain the fundamental mechanisms responsible for the distinct impacts of isolated fibers and intrinsic fibers on the gut microbiota of children with obesity.<sup>173</sup> In addition, DFs have a significant impact on the composition, diversity, abundance, and function of the gut microbiota in children with obesity, but the strength of the evidence remains insufficient. As shown in [Figure 1](#) and [Figure S1](#), there is a clear downward trend in Bacteroidetes, *Prevotella* and *Bacteroidetes* among children with obesity. Not surprisingly, following a high DF intake, all of these bacteria in children with obesity moved in a favorable direction ([Table 1](#)). There is still a large degree of uncertainty about the extent to which the effects attributed to DFs on obesity are mediated by the gut microbiota in children and the key species involved in the effects. However, current evidence still does not support the extent to which the effects of DFs on obesity are mediated by the gut microbiota in children and the key species involved in the effects.

However, there is no unanimous agreement on the recommended amounts of fiber for children with obesity, and there is a lack of evidence regarding practical advice on the appropriate type and quantity of DFs to be consumed. It is imperative to acknowledge that existing guidelines for DF consumption in children pertain to the overall intake of fibers and fail to sufficiently account for the origin and type of fibers.<sup>174</sup> This is important because various types and sources of fibers exhibit distinct physiological effects. For instance, the source (natural or synthetic, etc.), chemical composition (chain length, lignification, etc.), and physicochemical properties (solubility, viscosity, fermentability, etc.) of DFs affect not only the gut microbiota but also health outcomes.<sup>175,176</sup> Hence, in the formulation of a nutritional intervention program targeting children with obesity, it is imperative to take into account not only the overall daily consumption of DFs but also to acknowledge the origin and type of fiber duly. Furthermore,

additional research is required to investigate the physiological characteristics of the association between DFs and the health of children with obesity, as well as the optimal ratios of the different fiber sources.

Furthermore, the mechanism of action of DFs in regulating the gut microbiota to ameliorate childhood obesity has not been fully explored. We should be aware that the gut microbiota is an extraordinarily mutually dependent community in which waste products from one organism become nutrients for other organisms via substrate and metabolite cross-feeding. The cross-feeding behaviors of primary degraders and cross-feeders result in a wide range of substrates and metabolites to support microbiota diversity, allowing the gut to maintain a stable community.<sup>177</sup> For example, *in vitro* analysis revealed that the relationship between bifidobacteria and *F. prausnitzii*, in the presence of inulin-type fructans, could be commensal or competitive, and this relationship was dependent on the bifidobacterial strain and its capacity for prebiotics degradation.<sup>178</sup> More importantly, the gut microbiome profile of children with obesity is complex, with changes in the fungi, viruses, and archaea organisms in addition to bacterial homeostasis. To date, no studies have systematically reported the effects of DFs on the fungi, viruses and archaea organisms in the gut of children with obesity. Therefore, we should further our understanding of the complex relationships between fibers, gut microbiota, and childhood obesity, which can lead to the development of low-cost, safe, and efficacious “microbially-directed” dietary interventions.

#### 4. Future direction

Information on fiber-microbiome-obesity interactions in children needs to be further validated and refined. To be actionable, integration of the gut microbiome into the prevention of childhood obesity requires evidence of the causal contributions of the microbiome to obesity and the mechanistic physiological effects of fibers. Establishing the causal role of the gut microbiome in susceptibility to chronic diseases such as childhood obesity remains a challenge, which is complicated in nutrition research because the interactions between fibers, the gut microbiome,

and human health are complex and multidirectional. These complexities must be considered in the design of future nutrition research to elucidate what factors, including the gut microbiome, mediate the effects of fibers in childhood obesity. We suggest extending future studies through an experimental framework using three pillars that integrate the gut microbiome into stages of fiber nutrition research on children with obesity (Figure 3).

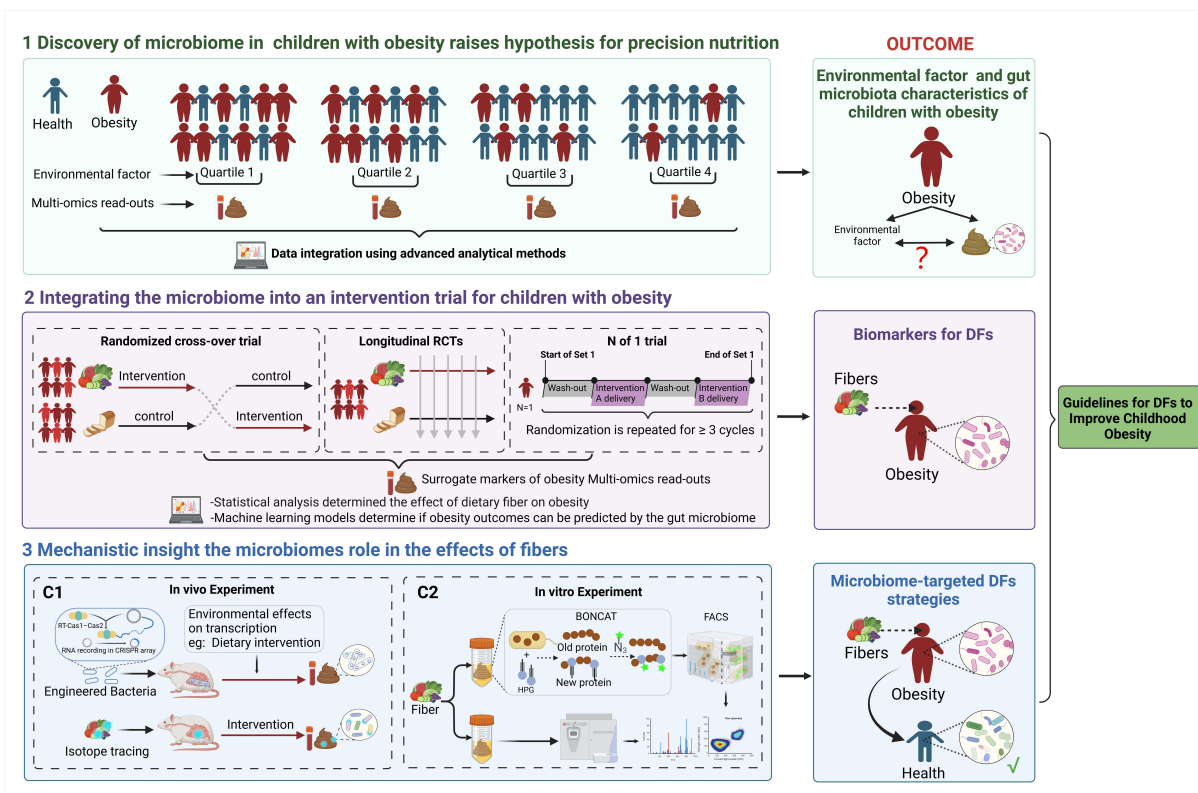
##### 4.1. Discovery of microbiome in children with obesity generates hypothesis for precision nutrition

Characterization of the gut microbiome may help in the prevention of childhood obesity and related metabolic diseases in the future. Furthermore, the application of multi-omics techniques (e.g., metagenomics, proteomics, metabolomics) and advanced analytical methods, such as machine learning-mediated analyses, has laid the groundwork for elucidating the underlying mechanisms and cause-and-effect relationships of biological pathways.<sup>179,180</sup> Recent studies have expanded on this framework by combining the microbiome and the epidemiology of childhood obesity to elucidate the microbiological profile of children with obesity.<sup>181</sup> Although such observational studies cannot establish causality, high-quality, large-scale cohort studies can help provide an evidence basis for the causes of childhood obesity. Integrating childhood obesity with microbiome epidemiology will facilitate the identification of links between the microbiome and childhood obesity and the characterization of the gut microbiota in children with obesity. The underlying mechanisms and biological plausibility of these interactions, as well as their value as diagnostic markers, can then be confirmed in experimental systems to confirm the system.

##### 4.2. Integrating the microbiome into nutritional intervention trials in childhood obesity

More clinical research using fiber interventions in childhood obesity, aimed at addressing the gut microbial perspective is needed, as the number of studies identified thus far is small, as there is a paucity of data available. Notably, although numerous epidemiological studies have demonstrated an association between DF intake and





**Figure 3.** Experimental framework of the fiber-microbiota-obesity study in children. (a) microbiome epidemiology can elucidate associations between the gut microbiome, environmental factors (e.g., diet, age, sex, etc.), and physiologic effects caused by environmental factors in children with obesity. This information does not establish causality, but can help to uncover gut microbiome characteristics in children with obesity and contribute to the evidence base for precision nutrition. (b) RCT can determine the causal relationship between DFs and childhood obesity and provide strong evidence of this. N of 1 employs the “individual” as the subject of the study, allowing for the development of personalized and precise nutritional requirements. Combined with microbiome-based multi-omics analysis, microbial biomarkers for precision nutrition strategies can be identified. However, highly personalized microbiomes and causal inference of their role in DF’s effects still pose challenges. (c) causal relationships and potential mechanisms can be further established in in vivo (animal models) and in vitro studies. Causal and mechanistic information provides additional evidence for the development of dietary guidelines and targeted nutritional strategies for DFs. BONCAT, bio-orthogonal non-canonical amino acid tagging; CRISPR, clustered, regularly interspaced, short palindromic repeats; DFs, dietary fibers; FACS, fluorescence-activated cell sorting; RCT, randomized controlled trial.

childhood obesity, there have been some inconsistent reports. In contrast, a clinical trial by Liber and Szajewska<sup>182</sup> showed that consumption of 8 g/d of oligofructose in otherwise healthy overweight or children with obesity aged 7–11 years and 15 g/d in children aged 12–18 years did not result in improvements in BMI or Z-score after 12 weeks compared with placebo control. There were also no improvements in the secondary outcomes of change in body weight and difference in absolute body fat or other metabolic outcomes. A systematic review and meta-analysis of the relationship between DFs and diabetes management found similar results.<sup>183</sup> These findings point to variation in the response of DFs to obesity-related metabolic

dysregulation in children, which is thought to be due to the high degree of individualization of gut microbes in children. The research of Deehan et al. supports this theory.<sup>184</sup> A DF intervention is likely to have no effect on an individual whose microbiome does not include keystone species or other genera that encode the enzymatic machinery to degrade those specific fiber types. This should thus be considered when designing future studies.

Randomized controlled trials are the gold standard for establishing causal effects. Optimally, feeding studies with single ingredient modifications using crossover designs with sufficient wash-out periods are ideal to assess not only the impact of DFs on gastrointestinal bacteria taxa, but also

microbial metabolites and other physiological measures of health such as body composition, blood cholesterol, glycemia, and inflammation. It is critical to include controls, which can eliminate inter-individual differences in individual-specific factors (e.g., microbiome, genetics, metabolite profiles, and baseline clinical measures).<sup>185</sup> Furthermore, for the highly individualized gut microbiome of children with obesity, a precision nutrition research design (N of 1 trial) can be used to screen for nutritional interventions that are appropriate for them, using themselves as a control, to achieve true, albeit difficult, precision nutrition.<sup>186</sup>

Studies should account for variation in other confounding variables such as demographics (age<sup>187</sup> and gender<sup>188</sup>) and lifestyle factors (such as habitual diet).<sup>189</sup> Obviously, in view of the complexity of the confounding variables, we need advanced statistical and modeling methods to carry out data analysis. For example, regression and correlation analyses can be applied to determine associations between fibers-induced changes in microbiome composition/functionality and clinical and mechanistic endpoints.<sup>190</sup> Machine-learning models can determine if fibers-induced physiological changes can be predicted by effects on the microbiome or biological processes in the host impacted by the microbiome.<sup>191</sup> Integration of the microbiome into human intervention trials could provide putative mechanistic explanations for the role of the microbiome in the effects of fibers on childhood obesity, as well as diagnostic microbiome-based biomarkers for precision nutrition strategies.

#### **4.3. Mechanistic insight the microbiomes role in the effects of fibers**

Human studies can be paired with animal models to determine whether the microbiome plays a causal role in the physiological effect, identify the causal components within the microbiome, and determine underlying mechanisms<sup>192</sup> (Figure 3 C1). The complex and dynamic transcriptional changes of the gut microbiota throughout time, transit, and perturbation in the gut of gnotobiotic animals are recorded by integrating RNA-derived spacers from the transcriptome of gut bacteria into plasmid DNA-encoded Clustered, regularly interspaced, short

palindromic repeats (CRISPR) arrays using F<sub>s</sub>RT-Cas1-Cas2<sup>193</sup> (Figure 3 C1). This scalable, noninvasive system for assessing intestinal function in vivo archives characteristics of microbial signatures of physiological or pathological states.<sup>177</sup> Transcriptome-scale recordings elucidate microbial responses to alterations in the intraluminal environment across nutrition and microbe-microbe interactions.<sup>177</sup> Furthermore, a method for tracing isotopes into bacterial-specific protein sequences has been developed that enables the mapping of nutrient routes in vivo and reveals how diet affects the makeup of the microbiome (Figure 3 C1). However, mechanistic studies in animals can be confounded as it is not possible to focus solely on the gut microbiota due to the presence of the host.<sup>194</sup> Noteworthy, in vitro gut fermentation models can be used to supplement human and animal studies and overcome some of the limitations of in vivo models<sup>195</sup> (Figure 3 C2). Researchers can use a variety of in vitro models in which single or multiple vessels are inoculated with fresh human feces or a defined microbial community. These vessels are operated under anaerobic conditions, and microbial communities are grown with temperature, pH, growth medium, and transit time set to mimic a specific intestinal segment. This tool can be used in combination with new technologies (e.g., Bio-orthogonal non-canonical amino acid tagging-Fluorescence-activated cell sorting [BONCAT-FACS],<sup>196</sup> Figure 3 C2) to test the direct effects of prebiotics and DFs on the gut microbiota of children with obesity in the absence of confounding factors. While both animal and in vitro models can mimic some aspects of the human gut microbiome, neither model can fully replicate all of the functions of the human gut. Despite their limitations, their microbiomes, if well controlled and combined with a multi-omics approach, have the capacity to complement human studies as they establish the mechanistic basis for the impact of fibers on children with obesity.

## **5. Conclusion**

The current evidences suggest that a higher intake of DFs may have a beneficial impact on metabolic health through modifications to the gut microbiota.<sup>175</sup> Personalized fiber intake

recommendations and improved obesity outcomes ought to be feasible with the integration of individual-level genetic and metabolic trait data, along with the development of microbial sequencing technologies and analytical methodologies. Microbiome research possesses the opportunity to furnish exhaustive insights into facets of nutrition strategy. However, the gut microbial signatures observed in children with obesity relative to normal-weight children remain unclear and require further research. Overall, in order to improve the convincing evidence supporting dietary guidelines for children, it is imperative that facets of childhood obesity prevention research incorporate gut microbial features.

## Abbreviations

|              |   |
|--------------|---|
| BAs          | Bile acids  |
| BONCAT-FACS  | Bio-orthogonal non-canonical amino acid tagging-Fluorescence-activated cell sorting |
| CRISPR       | Clustered, regularly interspaced, short palindromic repeats                         |
| DFs          | Dietary fibers  |
| FOS          | fructooligosaccharides  |
| FXR          | Farnesoid X receptor  |
| GPCR         | G protein-coupled receptors   |
| GLP-1        | glucagon-like peptide 1   |
| HDL          | high-density lipoprotein  |
| LPS          | Lipopolysaccharides   |
| PPAR         | peroxisome proliferator-activated receptor  |
| PYY          | peptide secretion YY  |
| RT-PCR       | Real-time PCR   |
| SCFAs        | short-chain fatty acids   |
| TNF $\alpha$ | tumor necrosis factor- $\alpha$   |
| UCP2         | uncoupling protein 2  |
| VLPs         | virus-like particles  |
| BMI          | body mass index   |

## Acknowledgments

Z.Y., J.L. and Z.Z. initiated the idea, scope, and outline of this review paper. Z.Y., C.C., and M.Y. conducted the systematic review and data extraction. Z.Y. and Z.Z. drafted the manuscript. Z.Y., J.L. and Z.Z. were responsible for conceptualizing the content of the figure. E.C.D., C.C., K.L.M., E.W., G.L., J.L. and Z.Z. critically reviewed and edited drafts of the manuscript. All authors approved the final version.




## Disclosure statement

No potential conflict of interest was reported by the author(s).

## Funding

This work was supported by the National Natural Science Foundation of China [No. 32202068] and the Natural Science Foundation of Fujian Province [No. 2022J01332].

## ORCID

Zhongmin Yang  <http://orcid.org/0000-0002-5570-2187>  
 Mingyue Yang  <http://orcid.org/0009-0007-5400-9878>  
 Edward C. Deehan  <http://orcid.org/0000-0001-7697-1418>  
 Chenxi Cai  <http://orcid.org/0009-0008-9318-1707>  
 Karen L. Madsen  <http://orcid.org/0000-0002-8636-0714>  
 Eytan Wine  <http://orcid.org/0000-0002-3458-4142>  
 Guiling Li  <http://orcid.org/0000-0002-0940-6010>  
 Jingwen Liu  <http://orcid.org/0000-0001-7231-6483>  
 Zhengxiao Zhang  <http://orcid.org/0000-0002-6977-4309>

## References

1. Bjerregaard LG, Jensen BW, Ängquist L, Osler M, Sørensen TIA, Baker JL. Change in overweight from childhood to early adulthood and risk of type 2 diabetes. *N Engl J Med.* 2018;378(14):1302–1312. doi:10.1056/NEJMoa1713231. PMID: 29617589.
2. Tricò D, Caprio S, Rosaria Umamo G, Pierpont B, Nouws J, Galderisi A, Kim G, Mata MM, Santoro N. Metabolic features of nonalcoholic fatty liver (NAFL) in obese adolescents: findings from a multiethnic cohort. *Hepatology.* 2018;68(4):1376–1390. doi:10.1002/hep.30035. PMID: 29665034.
3. Ortega FB, Lavie CJ, Blair SN. Obesity and cardiovascular disease. *Circ Res.* 2016;118(11):1752–1770. doi:10.1161/circresaha.115.306883. PMID: 27230640.
4. World Obesity Federation, World Obesity Atlas. 2023; <https://data.worldobesity.org/publications/?cat=19>.
5. Singh AS, Mulder C, Twisk JW, van Mechelen W, Chinapaw MJ. Tracking of childhood overweight into adulthood: a systematic review of the literature. *Obes Rev.* 2008;9(5):474–488. doi:10.1111/j.1467-789X.2008.00475.x. PMID: 18331423.
6. Quek YH, Tam WWS, Zhang MWB, Ho RCM. Exploring the association between childhood and adolescent obesity and depression: a meta-analysis. *Obes Rev.* 2017;18(7):742–754. doi:10.1111/obr.12535. PMID: 28401646.
7. Pereira LJ, Hinnig PF, Matsuo LH, Di Pietro PF, de Assis Maa, Vieira FGK. Association between lifestyle patterns and overweight and obesity in adolescents:

- a systematic review. *Br J Nutr.* 2022;1–45. doi:10.1017/s0007114522000228. PMID: 35086569.
8. Singh S, Ricardo-Silgado ML, Bielinski SJ, Acosta A. Pharmacogenomics of medication-induced weight gain and antiobesity medications. *Obesity (Silver Spring).* 2021;29(2):265–273. doi:10.1002/oby.23068. PMID: 33491309.
  9. Tzvetkov MV, Vormfelde SV, Balen D, Meineke I, Schmidt T, Sehr D, Sabolić I, Koepsell H, Brockmöller J. The effects of genetic polymorphisms in the organic cation transporters OCT1, OCT2, and OCT3 on the renal clearance of metformin. *Clin Pharmacol Ther.* 2009;86(3):299–306. doi:10.1038/clpt.2009.92. PMID: 19536068.
  10. Mohammadian Khonsari N, Khashayar P, Shahrestanaki E, Kelishadi R, Mohammadpoor Nami S, Heidari-Beni M, Esmaeili Abdar Z, Tabatabaei-Malazy O, Qorbani M. Normal weight obesity and cardiometabolic risk factors: a systematic review and meta-analysis. *Front Endocrinol (Lausanne).* 2022;13:857930. doi:10.3389/fendo.2022.857930. PMID: 35399938.
  11. Bouter KE, van Raalte Dh, Groen AK, Nieuwdorp M. Role of the gut microbiome in the pathogenesis of obesity and obesity-related metabolic dysfunction. *Gastroenterology.* 2017;152(7):1671–1678. doi:10.1053/j.gastro.2016.12.048. PMID: 28192102.
  12. Koboziev I, Reinoso Webb C, Furr KL, Grisham MB. Role of the enteric microbiota in intestinal homeostasis and inflammation. *Free Radic Biol Med.* 2014;68:122–133. doi:10.1016/j.freeradbiomed.2013.11.008. PMID: 24275541.
  13. Sender R, Fuchs S, Milo R. Revised estimates for the number of human and bacteria cells in the body. *PLoS Biol.* 2016;14(8):e1002533. doi:10.1371/journal.pbio.1002533. PMID: 27541692.
  14. Brahe LK, Astrup A, Larsen LH. Can we prevent obesity-related metabolic diseases by dietary modulation of the gut microbiota? *Adv Nutr.* 2016;7(1):90–101. doi:10.3945/an.115.010587. PMID: 26773017.
  15. de Wit Df, Hanssen NMJ, Wortelboer K, Herrema H, Rampanelli E, Nieuwdorp M. Evidence for the contribution of the gut microbiome to obesity and its reversal. *Sci Transl Med.* 2023;15(723):eadg2773. doi:10.1126/scitranslmed.adg2773. PMID: 37992156.
  16. Duranti S, Ferrario C, van Sinderen D, Ventura M, Turrone F. Obesity and microbiota: an example of an intricate relationship. *Genes Nutr.* 2017;12(1):18. doi:10.1186/s12263-017-0566-2. PMID: 28638490.
  17. Cunningham AL, Stephens JW, Harris DA. A review on gut microbiota: a central factor in the pathophysiology of obesity. *Lipids Health Dis.* 2021;20(1):65. doi:10.1186/s12944-021-01491-z. PMID: 34233682.
  18. Yatsunenko T, Rey FE, Manary MJ, Trehan I, Dominguez-Bello MG, Contreras M, Magris M, Hidalgo G, Baldassano RN, Anokhin AP, et al. Human gut microbiome viewed across age and geography. *Nature.* 2012;486(7402):222–227. doi:10.1038/nature11053. PMID: 22699611.
  19. Ronan V, Yeasin R, Claud EC. Childhood development and the microbiome—the intestinal microbiota in maintenance of health and development of disease during childhood development. *Gastroenterology.* 2021;160(2):495–506. doi:10.1053/j.gastro.2020.08.065. PMID: 33307032.
  20. Derrien M, Alvarez AS, de Vos WM. The gut microbiota in the first decade of life. *Trends Microbiol.* 2019;27(12):997–1010. doi:10.1016/j.tim.2019.08.001. PMID: 31474424.
  21. Wang Y, Salonen A, Jian C. Can prebiotics help tackle the childhood obesity epidemic? *Front Endocrinol (Lausanne).* 2023;14:1178155. doi:10.3389/fendo.2023.1178155. PMID: 37305030.
  22. Korczak R, Slavin JL. Definitions, regulations, and new frontiers for dietary fiber and whole grains. *Nutr Rev.* 2020;78(Supplement\_1):6–12. doi:10.1093/nutrit/nuz061. PMID: 32728750.
  23. Gibson GR, Hutkins R, Sanders ME, Prescott SL, Reimer RA, Salminen SJ, Scott K, Stanton C, Swanson KS, Cani PD, et al. Expert consensus document: the international scientific association for probiotics and prebiotics (ISAPP) consensus statement on the definition and scope of prebiotics. *Nat Rev Gastroenterol Hepatol.* 2017;14(8):491–502. doi:10.1038/nrgastro.2017.75. PMID: 28611480.
  24. Cronin P, Joyce SA, O’Toole PW, O’Connor EM. Dietary fibre modulates the gut microbiota. *Nutrients.* 2021;13(5):1655. doi:10.3390/nu13051655. PMID: 34068353.
  25. Tuncil YE, Thakkar RD, Arioglu-Tuncil S, Hamaker BR, Lindemann SR. Subtle variations in dietary-fiber fine structure differentially influence the composition and metabolic function of gut microbiota. *mSphere.* 2020;5(3). doi:10.1128/mSphere.00180-20. PMID: 32376698.
  26. Gill SK, Rossi M, Bajka B, Whelan K. Dietary fibre in gastrointestinal health and disease. *Nat Rev Gastroenterol Hepatol.* 2021;18(2):101–116. doi:10.1038/s41575-020-00375-4. PMID: 33208922.
  27. Zhu L, Baker SS, Gill C, Liu W, Alkhoury R, Baker RD, Gill SR. Characterization of gut microbiomes in non-alcoholic steatohepatitis (NASH) patients: a connection between endogenous alcohol and NASH. *Hepatology.* 2013;57(2):601–609. doi:10.1002/hep.26093. PMID: 23055155.
  28. Yuan X, Zhang Y, Lin X, Yang X, Chen R. Association of gut microbiota and glucose metabolism in children with disparate degrees of adiposity. *Pediatr Obes.* 2023;18(4):e13009. doi:10.1111/ijpo.13009. PMID: 36704910.
  29. Yuan X, Chen R, Zhang Y, Lin X, Yang X, McCormick KL. Gut microbiota of Chinese obese children and adolescents with and without insulin resistance. *Front Endocrinol (Lausanne).* 2021;12:636272. doi:10.3389/fendo.2021.636272. PMID: 33815293.

30. Yuan X, Chen R, McCormick KL, Zhang Y, Lin X, Yang X. The role of the gut microbiota on the metabolic status of obese children. *Microb Cell Fact*. 2021;20(1):53. doi:10.1186/s12934-021-01548-9. PMID: 33639944.
31. Wei Y, Liang J, Su Y, Wang J, Amakye WK, Pan J, Chu X, Ma B, Song Y, Li Y, et al. The associations of the gut microbiome composition and short-chain fatty acid concentrations with body fat distribution in children. *Clin Nutr*. 2021;40(5):3379–3390. doi:10.1016/j.clnu.2020.11.014. PMID: 33277072.
32. Wei J, Dai W, Pan X, Zhong Y, Xu N, Ye P, Wang J, Li J, Yang F, Luo J, et al. Identifying the novel gut microbial metabolite contributing to metabolic syndrome in children based on integrative analyses of microbiome-metabolome signatures. *Microbiol Spectr*. 2023;11(2):e0377122. doi:10.1128/spectrum.03771-22. PMID: 36794949.
33. Visuthranukul C, Sriswasdi S, Tepasorndech S, Joyjinda Y, Saengpanit P, Kwanbunbumpen T, Panichsillaphakit E, Uaariyapanichkul J, Chomtho S. Association of human intestinal microbiota with lifestyle activity, adiposity, and metabolic profiles in Thai children with obesity. *J Nutr Metab*. 2022;2022:1–14. doi:10.1155/2022/3029582. PMID: 35637874.
34. Vazquez-Moreno M, Perez-Herrera A, Locia-Morales D, Dizzel S, Meyre D, Stearns JC, Cruz M. Association of gut microbiome with fasting triglycerides, fasting insulin and obesity status in Mexican children. *Pediatr Obes*. 2021;16(5):e12748. doi:10.1111/ijpo.12748. PMID: 33191616.
35. Shin S, Cho KY. Altered gut microbiota and shift in bacteroidetes between young obese and normal-weight Korean children: a cross-sectional observational study. *Biomed Res Int*. 2020;2020:1–19. doi:10.1155/2020/6587136. PMID: 32908903.
36. Riva A, Borgo F, Lassandro C, Verduci E, Morace G, Borghi E, Berry D. Pediatric obesity is associated with an altered gut microbiota and discordant shifts in Firmicutes populations. *Environ Microbiol*. 2017;19(1):95–105. doi:10.1111/1462-2920.13463. PMID: 27450202.
37. Pierri L, Saggese P, Guercio Nuzio S, Troisi J, Di Stasi M, Poeta M, Savastano R, Marchese G, Tarallo R, Massa G, et al. Relations of gut liver axis components and gut microbiota in obese children with fatty liver: a pilot study. *Clin Res Hepatol Gastroenterol*. 2018;42(4):387–390. doi:10.1016/j.clinre.2018.03.015. PMID: 29773420.
38. Peng Y, Tan Q, Afhami S, Deehan EC, Liang S, Gantz M, Triador L, Madsen KL, Walter J, Tun HM, et al. The gut microbiota profile in children with prader-Willi Syndrome. *Genes (Basel)*. 2020;11(8):11. doi:10.3390/genes11080904. PMID: 32784572.
39. Pan X, Kaminga AC, Liu A, Wen SW, Luo M, Gut Microbiota LJ, Glucose. Gut microbiota, glucose, lipid, and water-electrolyte metabolism in children with nonalcoholic fatty liver disease. *Front Cell Infect Microbiol*. 2021;11:683743. doi:10.3389/fcimb.2021.683743. PMID: 34778099.
40. Nirmalkar K, Murugesan S, Pizano-Zárate ML, Villalobos-Flores LE, García-González C, Morales-Hernández RM, Nuñez-Hernández JA, Hernández-Quiroz F, Romero-Figueroa MDS, Hernández-Guerrero C, et al. Gut microbiota and endothelial dysfunction markers in obese Mexican children and adolescents. *Nutrients*. 2018;10(12):2009. doi:10.3390/nu10122009. PMID: 30572569.
41. Murugesan S, Ulloa-Martínez M, Martínez-Rojano H, Galván-Rodríguez FM, Miranda-Brito C, Romano MC, Piña-Escobedo A, Pizano-Zárate ML, Hoyo-Vadillo C, García-Mena J. Study of the diversity and short-chain fatty acids production by the bacterial community in overweight and obese Mexican children. *Eur J Clin Microbiol Infect Dis*. 2015;34(7):1337–1346. doi:10.1007/s10096-015-2355-4. PMID: 25761741.
42. Monga Kravetz A, Testerman T, Galuppo B, Graf J, Pierpont B, Siebel S, Feinn R, Santoro N. Effect of gut microbiota and PNPLA3 rs738409 variant on nonalcoholic fatty liver disease (NAFLD) in obese youth. *J Clin Endocrinol Metab*. 2020;105(10):e3575–85. doi:10.1210/clinem/dgaa382. PMID: 32561908.
43. Méndez-Salazar EO, Ortiz-López MG, Granados-Silvestre M, Palacios-González B, Menjivar M. Altered gut microbiota and compositional changes in Firmicutes and Proteobacteria in Mexican undernourished and obese children. *Front Microbiol*. 2018;9:2494. doi:10.3389/fmicb.2018.02494. PMID: 30386323.
44. Ma T, Wu Z, Lin J, Shan C, Abasijiang A, Zhao J. Characterization of the oral and gut microbiome in children with obesity aged 3 to 5 years. *Front Cell Infect Microbiol*. 2023;13:1102650. doi:10.3389/fcimb.2023.1102650. PMID: 37065198.
45. López-Contreras BE, Morán-Ramos S, Villarruel-Vázquez R, Macías-Kauffer L, Villamil-Ramírez H, León-Mimila P, Vega-Badillo J, Sánchez-Muñoz F, Llanos-Moreno LE, Canizalez-Román A, et al. Composition of gut microbiota in obese and normal-weight Mexican school-age children and its association with metabolic traits. *Pediatr Obes*. 2018;13(6):381–388. doi:10.1111/ijpo.12262. PMID: 29388394.
46. Karvonen AM, Sordillo JE, Gold DR, Bacharier LB, O'Connor GT, Zeiger RS, Beigelman A, Weiss ST, Litonjua AA. Gut microbiota and overweight in 3-year old children. *Int J Obes (Lond)*. 2019;43(4):713–723. doi:10.1038/s41366-018-0290-z. PMID: 30568265.
47. Jiang LF, Wang YY, Peng H, Li R, Zhang F, Wang N, Shao QW, Jiang Q. Association between obesity with the diversity and genus of gut microbiota in school-aged children. *Zhonghua Liu Xing Bing Xue Za Zhi*. 2022;43(2):260–268. doi:10.3760/cma.j.cn112338-20210617-00478. PMID: 35184494.

48. Jaimes JD, A S, Hurych J, Cinek O, Nichols B, Vodolánová L, Černý K, Havlík J, Ishaq SL. Stool metabolome-microbiota evaluation among children and adolescents with obesity, overweight, and normal-weight using 1H NMR and 16S rRNA gene profiling. *PLOS ONE*. 2021;16(3):e0247378. doi:10.1371/journal.pone.0247378. PMID: 33765008.
49. Hu HJ, Park SG, Jang HB, Choi MK, Park KH, Kang JH, Park SI, Lee HJ, Cho SH. Obesity alters the microbial community profile in Korean adolescents. *PLOS ONE*. 2015;10(7):e0134333. doi:10.1371/journal.pone.0134333. PMID: 26230509.
50. Gyarmati P, Song Y, Dotimas J, Yoshiba G, Christison A. Cross-sectional comparisons of gut microbiome and short-chain fatty acid levels among children with varied weight classifications. *Pediatr Obes*. 2021;16(6):e12750. doi:10.1111/ijpo.12750. PMID: 33174684.
51. Grigorova EV, Belkova NL, Nemchenko UM, Klimenko ES, Pogodina AV, Romanitsa AI, Novikova EA, Rychkova LV. Metasequencing of V3-V4 variable regions of 16S rRNA gene in opportunistic microbiota and gut biocenosis in obese adolescents. *Bull Exp Biol Med*. 2021;170(3):321–325. doi:10.1007/s10517-021-05060-3. PMID: 33452983.
52. Gao XF, Wu BB, Pan YL, Zhou SM, Zhang M, You YH, Cai YP, Liang Y. Gut microbiome biomarkers in adolescent obesity: a regional study. *Health Inf Sci Syst*. 2023;11(1):37. doi:10.1007/s13755-023-00236-9. PMID: 37602197.
53. Gao X, Jia R, Xie L, Kuang L, Feng L, Wan C. A study of the correlation between obesity and intestinal flora in school-age children. *Sci Rep*. 2018;8(1):14511. doi:10.1038/s41598-018-32730-6. PMID: 30267022.
54. Gallardo-Becerra L, Cornejo-Granados F, García-López R, Valdez-Lara A, Bikel S, Canizales-Quinteros S, López-Contreras BE, Mendoza-Vargas A, Nielsen H, Ochoa-Leyva A. Metatranscriptomic analysis to define the secrebiome, and 16S rRNA profiling of the gut microbiome in obesity and metabolic syndrome of Mexican children. *Microb Cell Fact*. 2020;19(1):61. doi:10.1186/s12934-020-01319-y. PMID: 32143621.
55. Ferrer M, Ruiz A, Lanza F, Haange SB, Oberbach A, Till H, Bargiela R, Campoy C, Segura MT, Richter M, et al. Microbiota from the distal guts of lean and obese adolescents exhibit partial functional redundancy besides clear differences in community structure. *Environ Microbiol*. 2013;15(1):211–226. doi:10.1111/j.1462-2920.2012.02845.x. PMID: 22891823.
56. Del Chierico F, Manco M, Gardini S, Guarrasi V, Russo A, Bianchi M, Tortosa V, Quagliariello A, Shashaj B, Fintini D, et al. Fecal microbiota signatures of insulin resistance, inflammation, and metabolic syndrome in youth with obesity: a pilot study. *Acta Diabetol*. 2021;58(8):1009–1022. doi:10.1007/s00592-020-01669-4. PMID: 33754165.
57. Del Chierico F, Abbatini F, Russo A, Quagliariello A, Reddel S, Capoccia D, Caccamo R, Ginanni Corradini S, Nobili V, De Peppo F, et al. Gut microbiota markers in obese adolescent and adult patients: age-dependent differential patterns. *Front Microbiol*. 2018;9:1210. doi:10.3389/fmicb.2018.01210. PMID: 29922272.
58. Da Silva CC, Monteil MA, Davis EM. Overweight and obesity in children are associated with an abundance of Firmicutes and reduction of bifidobacterium in their gastrointestinal microbiota. *Child Obes*. 2020;16(3):204–210. doi:10.1089/chi.2019.0280. PMID: 31934770.
59. Cho KY. Lifestyle modifications result in alterations in the gut microbiota in obese children. *BMC Microbiol*. 2021;21(1):10. doi:10.1186/s12866-020-02002-3. PMID: 33407104.
60. Chen X, Zhang D, Sun H, Jiang F, Shen Y, Wei P, Shen X. Characterization of the gut microbiota in Chinese children with overweight and obesity using 16S rRNA gene sequencing. *PeerJ*. 2021;9:e11439. doi:10.7717/peerj.11439. PMID: 34164233.
61. Chen X, Sun H, Jiang F, Shen Y, Li X, Hu X, Shen X, Wei P. Alteration of the gut microbiota associated with childhood obesity by 16S rRNA gene sequencing. *PeerJ*. 2020;8:e8317. doi:10.7717/peerj.8317. PMID: 31976177.
62. Benítez-Páez A, Gómez Del Pugar EM, López-Almela I, Moya-Pérez Á, Codoñer-Franch P, Sanz Y, Turnbaugh PJ. Depletion of Blautia Species in the microbiota of obese children relates to intestinal inflammation and metabolic phenotype worsening. *mSystems*. 2020;5(2). doi:10.1128/mSystems.00857-19. PMID: 32209719.
63. Balakrishnan B, Selvaraju V, Chen J, Ayine P, Yang L, Babu JR, Geetha T, Taneja V. Ethnic variability associating gut and oral microbiome with obesity in children. *Gut Microbes*. 2021;13(1):1–15. doi:10.1080/19490976.2021.1882926. PMID: 33596768.
64. Jf SM, Qiong W. Analysis of intestinal microecological status and influencing factors in obese children aged 3-6 years. *China Maternal And Child Health*. 2022;37:2981–2984. doi:10.19829/j.zgfybj.issn.1001-4411.2022.16.021.
65. Chunfeng T, Long G, Yufan G, Kai L, Yan B. A multi-omics study of intestinal flora and metabolism in obese Mongolian children. *Genomics And Appl Biol*. 2023;42:542–551. doi:10.13417/j.gab.042.000542.
66. Jing L, Yuanyuan L, Chaonan F, Xu D, Kemin Q. Changes of intestinal flora in preschool obese children. *Chinese journal of child health*. 2015;23:388–390.
67. Alcazar M, Escribano J, Ferré N, Closa-Monasterolo R, Selma-Royo M, Feliu A, Castillejo G, Luque V. Gut microbiota is associated with metabolic health in children with obesity. *Clin Nutr*. 2022;41(8):1680–1688. doi:10.1016/j.clnu.2022.06.007. PMID: 35777107.

68. Hou Y-P, He Q-Q, Ouyang H-M, Peng H-S, Wang Q, Li J, Lv X-F, Zheng Y-N, Li S-C, Liu H-L, et al. Human gut microbiota associated with obesity in Chinese children and adolescents. *Biomed Res Int*. 2017;2017:7585989. doi:10.1155/2017/7585989.
69. Grunec L, Gentekaki E, Kespechara K, Denny J, Sharpton TJ, Marriott LK, Shannon J, Popluechai S. The fecal microbiota of Thai school-aged children associated with demographic factors and diet. *PeerJ*. 2022;10:e13325. doi:10.7717/peerj.13325. PMID: 35469202.
70. Xu P, Li M, Zhang J, Zhang T. Correlation of intestinal microbiota with overweight and obesity in Kazakh school children. *BMC Microbiol*. 2012;12(1):283. doi:10.1186/1471-2180-12-283. PMID: 23190705.
71. Orbe-Orihuela YC, Lagunas-Martínez A, Bahena-Román M, Madrid-Marina V, Torres-Poveda K, Flores-Alfaro E, Méndez-Padrón A, Díaz-Benítez CE, Peralta-Zaragoza O, Antúnez-Ortiz D, et al. High relative abundance of firmicutes and increased tnf- $\alpha$  levels correlate with obesity in children. *Salud Publica Mex*. 2018;60(1, ene-feb):5–11. doi:10.21149/8133. PMID: 29689651.
72. Mousavi SH, Mehrara S, Barzegari A, Ostadrahimi A. Correlation of gut microbiota profile with body mass index among school age children. *Iran Red Crescent Med J*. 2018;20(4). doi:10.5812/ircmj.58049.
73. Karlsson CL, Onnerfält J, Xu J, Molin G, Ahrné S, Thorngren-Jerneck K. The microbiota of the gut in preschool children with normal and excessive body weight. *Obesity (Silver Spring)*. 2012;20(11):2257–2261. doi:10.1038/oby.2012.110. PMID: 22546742.
74. Goffredo M, Mass K, Parks EJ, Wagner DA, McClure EA, Graf J, Savoye M, Pierpont B, Cline G, Santoro N. Role of gut microbiota and short chain fatty acids in modulating energy harvest and fat partitioning in youth. *J Clin Endocrinol Metab*. 2016;101(11):4367–4376. doi:10.1210/jc.2016-1797. PMID: 27648960.
75. Bervoets L, Van Hoorenbeek K, Kortleven I, Van Noten C, Hens N, Vael C, Goossens H, Desager KN, Vankerckhoven V. Differences in gut microbiota composition between obese and lean children: a cross-sectional study. *Gut Pathog*. 2013;5(1):10. doi:10.1186/1757-4749-5-10. PMID: 23631345.
76. Balamurugan R, George G, Kabeerdoss J, Hepsiba J, Chandragunasekaran AM, Ramakrishna BS. Quantitative differences in intestinal *Faecalibacterium prausnitzii* in obese Indian children. *Br J Nutr*. 2010;103(3):335–338. doi:10.1017/s0007114509992182. PMID: 19849869.
77. Jy DZ, Junmei Y. Survey on the current status of overweight and obesity in preschool children and analysis of the results of intestinal flora testing. *China Convalescent Med*. 2019;28:1226–1228. doi:10.13517/j.cnki.ccm.2019.11.045.
78. Maya-Lucas O, Murugesan S, Nirmalkar K, Alcaraz LD, Hoyo-Vadillo C, Pizano-Zárate ML, García-Mena J. The gut microbiome of Mexican children affected by obesity. *Anaerobe*. 2019;55:11–23. doi:10.1016/j.anaerobe.2018.10.009. PMID: 30366118.
79. Zhao Y, Zhou J, Liu J, Wang Z, Chen M, Zhou S. Metagenome of gut microbiota of children with non-alcoholic fatty liver disease. *Front Pediatr*. 2019;7:518. doi:10.3389/fped.2019.00518. PMID: 31921729.
80. Testerman T, Li Z, Galuppo B, Graf J, Santoro N. Insights from shotgun metagenomics into bacterial species and metabolic pathways associated with NAFLD in obese youth. *Hepatol Commun*. 2022;6(8):1962–1974. doi:10.1002/hep4.1944. PMID: 35344283.
81. Orsso CE, Peng Y, Deehan EC, Tan Q, Field CJ, Madsen KL, Walter J, Prado CM, Tun HM, Haqq AM. Composition and functions of the gut microbiome in pediatric obesity: relationships with markers of insulin resistance. *Microorganisms*. 2021;9(7):1490. doi:10.3390/microorganisms9071490. PMID: 34361925.
82. Murga-Garrido SM, Ulloa-Pérez EJ, Díaz-Benítez CE, Orbe-Orihuela YC, Cornejo-Granados F, Ochoa-Leyva A, Sanchez-Flores A, Cruz M, Castañeda-Márquez AC, Plett-Torres T, et al. Virulence factors of the gut microbiome are associated with BMI and Metabolic blood parameters in children with obesity. *Microbiol Spectr*. 2023;11(2):e0338222. doi:10.1128/spectrum.03382-22. PMID: 36786619.
83. Michail S, Lin M, Frey MR, Fanter R, Paliy O, Hilbush B, Reo NV. Altered gut microbial energy and metabolism in children with non-alcoholic fatty liver disease. *FEMS Microbiol Ecol*. 2015;91(2):1–9. doi:10.1093/femsec/fiu002. PMID: 25764541.
84. Abdallah Ismail N, Ragab SH, Abd Elbaky A, Shoeib AR, Alhosary Y, Fekry D. Frequency of Firmicutes and Bacteroidetes in gut microbiota in obese and normal weight Egyptian children and adults. *Arch Med Sci*. 2011;7:501–507. doi:10.5114/aoms.2011.23418. PMID: 22295035.
85. Barczyńska R, Litwin M, Sliżewska K, Szalecki M, Berdowska A, Bandurska K, Libudzisz Z, Kapuśniak J. Bacterial microbiota and fatty acids in the faeces of overweight and obese children. *Pol J Microbiol*. 2018;67(3):339–345. doi:10.21307/pjm-2018-041. PMID: 30451451.
86. Nemchenko U, Belkova N, Pogodina A, Romanitsa A, Novikova E, Klimenko E, Grigorova E, Kungurtseva E, Butakova K, Rychkova L. Features of the composition of bifidoflora in the intestinal microbiome of obese adolescents. *Mol Genet Microbiol Virol*. 2021;36(3):144–151. doi:10.3103/s0891416821030046.
87. Ignacio A, Fernandes MR, Rodrigues VA, Groppo FC, Cardoso AL, Avila-Campos MJ, Nakano V. Correlation between body mass index and faecal microbiota from children. *Clin Microbiol Infect*. 2016;22(3):258.e1–8. doi:10.1016/j.cmi.2015.10.031. PMID: 26551842.

88. Borgo F, Verduci E, Riva A, Lassandro C, Riva E, Morace G, Borghi E. Relative abundance in bacterial and fungal gut microbes in obese children: a case control study. *Child Obes.* **2017**;13(1):78–84. doi:10.1089/chi.2015.0194. PMID: 27007700.
89. Luoto R, Kalliomäki M, Laitinen K, Delzenne NM, Cani PD, Salminen S, Isolauri E. Initial dietary and microbiological environments deviate in normal-weight compared to overweight children at 10 years of age. *J Pediatr Gastroenterol Nutr.* **2011**;52(1):90–95. doi:10.1097/MPG.0b013e3181f3457f. PMID: 21150648.
90. Shade A. Diversity is the question, not the answer. *The ISME J.* **2017**;11(1):1–6. doi:10.1038/ismej.2016.118. PMID: 27636395.
91. Crovesy L, Masterson D, Rosado EL. Profile of the gut microbiota of adults with obesity: a systematic review. *Eur J Clin Nutr.* **2020**;74(9):1251–1262. doi:10.1038/s41430-020-0607-6. PMID: 32231226.
92. Pinart M, Dötsch A, Schlicht K, Laudes M, Bouwman J, Forslund SK, Pischon T, Nimptsch K. Gut microbiome composition in obese and non-obese persons: a systematic review and meta-analysis. *Nutrients.* **2021**;14(1):12. doi:10.3390/nu14010012. PMID: 35010887.
93. Depommier C, Everard A, Druart C, Plovier H, Van Hul M, Vieira-Silva S, Falony G, Raes J, Maiter D, Delzenne NM, et al. Supplementation with *Akkermansia muciniphila* in overweight and obese human volunteers: a proof-of-concept exploratory study. *Nat Med.* **2019**;25(7):1096–1103. doi:10.1038/s41591-019-0495-2. PMID: 31263284.
94. Hold GL, Schwiertz A, Aminov RI, Blaut M, Flint HJ. Oligonucleotide probes that detect quantitatively significant groups of butyrate-producing bacteria in human feces. *Appl Environ Microbiol.* **2003**;69(7):4320–4324. doi:10.1128/aem.69.7.4320-4324.2003. PMID: 12839823.
95. Maioli TU, Borrás-Nogues E, Torres L, Barbosa SC, Martins VD, Langella P, Azevedo VA, Chatel JM. Possible benefits of faecalibacterium prausnitzii for obesity-associated gut disorders. *Front Pharmacol.* **2021**;12:740636. doi:10.3389/fphar.2021.740636. PMID: 34925006.
96. Shuai M, Fu Y, Zhong HL, Gou W, Jiang Z, Liang Y, Miao Z, Xu JJ, Huynh T, Wahlqvist ML, et al. Mapping the human gut mycobiome in middle-aged and elderly adults: multiomics insights and implications for host metabolic health. *Gut.* **2022**;71(9):1812–1820. doi:10.1136/gutjnl-2021-326298. PMID: 35017200.
97. Kaplan RC, Wang Z, Usyk M, Sotres-Alvarez D, Daviğlus ML, Schneiderman N, Talavera GA, Gellman MD, Thyagarajan B, Moon JY, et al. Gut microbiome composition in the Hispanic Community Health Study/Study of Latinos is shaped by geographic relocation, environmental factors, and obesity. *Genome Biol.* **2019**;20(1):219. doi:10.1186/s13059-019-1831-z. PMID: 31672155.
98. Huseyin CE, O'Toole PW, Cotter PD, Scanlan PD. Forgotten fungi—the gut mycobiome in human health and disease. *FEMS Microbiol Rev.* **2017**;41(4):479–511. doi:10.1093/femsre/fuw047. PMID: 28430946.
99. Mba IE, Nweze EI. Mechanism of *Candida* pathogenesis: revisiting the vital drivers. *Eur J Clin Microbiol Infect Dis.* **2020**;39(10):1797–1819. doi:10.1007/s10096-020-03912-w. PMID: 32372128.
100. Polke M, Hube B, Jacobsen ID. *Candida* survival strategies. *Adv Appl Microbiol.* **2015**;91:139–235. doi:10.1016/bs.aambs.2014.12.002. PMID: 25911234.
101. Sun S, Sun L, Wang K, Qiao S, Zhao X, Hu X, Chen W, Zhang S, Li H, Dai H, et al. The gut commensal fungus, *Candida parapsilosis*, promotes high fat-diet induced obesity in mice. *Commun Biol.* **2021**;4(1):1220. doi:10.1038/s42003-021-02753-3. PMID: 34697386.
102. Mar Rodríguez M, Pérez D, Javier Chaves F, Esteve E, Marin-García P, Xifra G, Vendrell J, Jové M, Pamplona R, Ricart W, et al. Obesity changes the human gut mycobiome. *Sci Rep.* **2015**;5(1):14600. doi:10.1038/srep14600. PMID: 26455903.
103. Hallen-Adams HE, Kachman SD, Kim J, Legge RM, Martínez I. Fungi inhabiting the healthy human gastrointestinal tract: a diverse and dynamic community. *Fungal Ecol.* **2015**;15:9–17. doi:10.1016/j.funeco.2015.01.006.
104. Wu L, Zeng T, Deligios M, Milanese L, Langille MGI, Zinellu A, Rubino S, Carru C, Kelvin DJ. Age-related variation of bacterial and fungal communities in different body habitats across the young, elderly, and centenarians in Sardinia. *mSphere.* **2020**;5(1). doi:10.1128/mSphere.00558-19. PMID: 32102941.
105. García-Gamboa R, Kirchmayr MR, Gradilla-Hernández MS, Pérez-Brocal V, Moya A, González-Avila M. The intestinal mycobiota and its relationship with overweight, obesity and nutritional aspects. *J Hum Nutr Diet.* **2021**;34(4):645–655. doi:10.1111/jhn.12864. PMID: 33586805.
106. Sokol H, Leducq V, Aschard H, Pham HP, Jegou S, Landman C, Cohen D, Liguori G, Bourrier A, Nion-Larmurier I, et al. Fungal microbiota dysbiosis in IBD. *Gut.* **2017**;66(6):1039–1048. doi:10.1136/gutjnl-2015-310746. PMID: 26843508.
107. Hills RD Jr., Pontefract BA, Mishcon HR, Black CA, Sutton SC, Theberge CR. Gut microbiome: profound implications for diet and disease. *Nutrients.* **2019**;11(7):11. doi:10.3390/nu11071613. PMID: 31315227.
108. Lambooi JM, Hoogenkamp MA, Brandt BW, Janus MM, Krom BP. Fungal mitochondrial oxygen consumption induces the growth of strict anaerobic bacteria. *Fungal Genet Biol.* **2017**;109:1–6. doi:10.1016/j.fgb.2017.10.001. PMID: 28989089.
109. van Leeuwen Pt, van der Peet Jm, Bikker FJ, Hoogenkamp MA, Oliveira Paiva AM, Kostidis S, Mayboroda OA, Smits WK, Krom BP. Interspecies



- interactions between *Clostridium difficile* and *Candida albicans*. *mSphere*. 2016;1(6). doi:10.1128/mSphere.00187-16. PMID: 27840850.
110. Shabana SS, Irfan U. The gut microbiota and its potential role in obesity. *Future Microbiol*. 2018;13:589–603. doi:10.2217/fmb-2017-0179. PMID: 29533087.
  111. Zhang T, Breitbart M, Lee WH, Run JQ, Wei CL, Soh SW, Hibberd ML, Liu ET, Rohwer F, Ruan Y. RNA viral community in human feces: prevalence of plant pathogenic viruses. *PLoS Biol*. 2006;4(1):e3. doi:10.1371/journal.pbio.0040003. PMID: 16336043.
  112. Yadav H, Jain S, Nagpal R, Marotta F. Increased fecal viral content associated with obesity in mice. *World J Diabetes*. 2016;7(15):316–320. doi:10.4239/wjd.v7.i15.316. PMID: 27555892.
  113. Bikel S, López-Leal G, Cornejo-Granados F, Gallardo-Becerra L, García-López R, Sánchez F, Equihua-Medina E, Ochoa-Romo JP, López-Contreras BE, Canizales-Quinteros S, et al. Gut dsDNA virome shows diversity and richness alterations associated with childhood obesity and metabolic syndrome. *iScience*. 2021;24(8):102900. doi:10.1016/j.isci.2021.102900. PMID: 34409269.
  114. Bikel S, López-Leal G, Cornejo-Granados F, Gallardo-Becerra L, Sánchez F, Equihua-Medina E, Ochoa-Romo JP, López-Contreras BE, Canizales-Quinteros S, Leyva AO. Gut phageome analysis reveals disease-specific hallmarks in childhood obesity. *bioRxiv*. 2020:2020.07.29.227637. doi: 10.1101/2020.07.29.227637.
  115. Cervantes-Echeverría M, Gallardo-Becerra L, Cornejo-Granados F, Ochoa-Leyva A. The two-faced role of crAssphage subfamilies in obesity and metabolic syndrome: between good and evil. *Genes (Basel)*. 2023;14(1):139. doi:10.3390/genes14010139. PMID: 36672880.
  116. Dutilh BE, Cassman N, McNair K, Sanchez SE, Silva GG, Boling L, Barr JJ, Speth DR, Seguritan V, Aziz RK, et al. A highly abundant bacteriophage discovered in the unknown sequences of human faecal metagenomes. *Nat Commun*. 2014;5(1):4498. doi:10.1038/ncomms5498. PMID: 25058116.
  117. Shen J, Zhang J, Mo L, Li Y, Li Y, Li C, Kuang X, Tao Z, Qu Z, Wu L, et al. Large-scale phage cultivation for commensal human gut bacteria. *Cell Host & Microbe*. 2023;31(4):665–77.e7. doi:10.1016/j.chom.2023.03.013. PMID: 37054680.
  118. Ott SJ, Waetzig GH, Rehman A, Moltzau-Anderson J, Bharti R, Grasis JA, Cassidy L, Tholey A, Fickenscher H, Seegert D, et al. Efficacy of sterile fecal filtrate transfer for treating patients with *clostridium difficile* infection. *Gastroenterology*. 2017;152(4):799–811.e7. doi:10.1053/j.gastro.2016.11.010. PMID: 27866880.
  119. Miller TL, Wolin MJ, Conway de Macario E, Macario AJ. Isolation of *methanobrevibacter smithii* from human feces. *Appl Environ Microbiol*. 1982;43(1):227–232. doi:10.1128/aem.43.1.227-232.1982. PMID: 6798932.
  120. Miller TL, Wolin MJ. *Methanosphaera stadtmaniae* gen. nov. sp. nov.: a species that forms methane by reducing methanol with hydrogen. *Arch Microbiol*. 1985;141(2):116–122. doi:10.1007/bf00423270. PMID: 3994486.
  121. Dridi B, Fardeau ML, Ollivier B, Raoult D, Drancourt M. *Methanomassiliicoccus luminyensis* gen. nov. sp. nov. a methanogenic archaeon isolated from human faeces. *Int J Syst Evol Microbiol*. 2012;62(Pt\_8):1902–1907. doi:10.1099/ijs.0.033712-0. PMID: 22859731.
  122. Samuel BS, Gordon JI. A humanized gnotobiotic mouse model of host–archaeal–bacterial mutualism. *Proc Natl Acad Sci USA*. 2006;103(26):10011–10016. doi:10.1073/pnas.0602187103. PMID: 16782812.
  123. Samuel BS, Hansen EE, Manchester JK, Coutinho PM, Henrissat B, Fulton R, Latreille P, Kim K, Wilson RK, Gordon JI. Genomic and metabolic adaptations of *methanobrevibacter smithii* to the human gut. *Proc Natl Acad Sci USA*. 2007;104(25):10643–10648. doi:10.1073/pnas.0704189104. PMID: 17563350.
  124. Gregory AC, Zablocki O, Zayed AA, Howell A, Bolduc B, Sullivan MB. The gut virome database reveals age-dependent patterns of virome diversity in the human gut. *Cell Host Microbe*. 2020;28(5):724–40.e8. doi:10.1016/j.chom.2020.08.003. PMID: 32841606.
  125. Le Poul E, Loison C, Struyf S, Springael JY, Lannoy V, Decobecq ME, Brezillon S, Dupriez V, Vassart G, Van Damme J, et al. Functional characterization of human receptors for short chain fatty acids and their role in polymorphonuclear cell activation. *J Biol Chem*. 2003;278(28):25481–25489. doi:10.1074/jbc.M301403200. PMID: 12711604.
  126. Husted AS, Trauelsen M, Rudenko O, Hjorth SA, Schwartz TW. GPCR-Mediated signaling of metabolites. *Cell Metab*. 2017;25(4):777–796. doi:10.1016/j.cmet.2017.03.008. PMID: 28380372.
  127. Ahmed K, Tunaru S, Offermanns S. GPR109A, GPR109B and GPR81, a family of hydroxy-carboxylic acid receptors. *Trends Pharmacol Sci*. 2009;30(11):557–562. doi:10.1016/j.tips.2009.09.001. PMID: 19837462.
  128. Lin HV, Frassetto A, Kowalik EJ Jr., Nawrocki AR, Lu MM, Kosinski JR, Hubert JA, Szeto D, Yao X, Forrest G, et al. Butyrate and propionate protect against diet-induced obesity and regulate gut hormones via free fatty acid receptor 3-independent mechanisms. *PLoS One*. 2012;7(4):e35240. doi:10.1371/journal.pone.0035240. PMID: 22506074.
  129. Coppola S, Avagliano C, Calignano A, Berni Canani R. The protective role of butyrate against obesity and obesity-related diseases. *Molecules*. 2021;26(3):682. doi:10.3390/molecules26030682. PMID: 33525625.
  130. den Besten G, van Eunen K, Groen AK, Venema K, Reijngoud DJ, Bakker BM. The role of short-chain fatty acids in the interplay between diet, gut microbiota, and host energy metabolism. *J Lipid Res*. 2013;54

- (9):2325–2340. doi:10.1194/jlr.R036012. PMID: 23821742.
131. Delzenne NM, Neyrinck AM, Bäckhed F, Cani PD. Targeting gut microbiota in obesity: effects of prebiotics and probiotics. *Nat Rev Endocrinol.* 2011;7(11):639–646. doi:10.1038/nrendo.2011.126. PMID: 21826100.
132. Usuda H, Okamoto T, Wada K. Leaky gut: effect of dietary fiber and fats on microbiome and intestinal barrier. *Int J Mol Sci.* 2021;22(14):7613. doi:10.3390/ijms22147613. PMID: 34299233.
133. Boutagy NE, McMillan RP, Frisard MI, Hulver MW. Metabolic endotoxemia with obesity: Is it real and is it relevant? *Biochimie.* 2016;124:11–20. doi:10.1016/j.biochi.2015.06.020. PMID: 26133659.
134. Akhtar M, Chen Y, Ma Z, Zhang X, Shi D, Khan JA, Liu H. Gut microbiota-derived short chain fatty acids are potential mediators in gut inflammation. *Anim Nutr.* 2022;8:350–360. doi:10.1016/j.aninu.2021.11.005. PMID: 35510031.
135. Wrzosek L, Miquel S, Noordine ML, Bouet S, Joncquel Chevalier-Curt M, Robert V, Philippe C, Bridonneau C, Cherbuy C, Robbe-Masselot C, et al. Bacteroides thetaiotaomicron and Faecalibacterium prausnitzii influence the production of mucus glycans and the development of goblet cells in the colonic epithelium of a gnotobiotic model rodent. *BMC Biol.* 2013;11(1):61. doi:10.1186/1741-7007-11-61. PMID: 23692866.
136. Byndloss MX, Olsan EE, Rivera-Chávez F, Tiffany CR, Cevallos SA, Lokken KL, Torres TP, Byndloss AJ, Faber F, Gao Y, et al. Microbiota-activated ppar- $\gamma$  signaling inhibits dysbiotic Enterobacteriaceae expansion. *Science.* 2017;357(6351):570–575. doi:10.1126/science.aam9949. PMID: 28798125.
137. Wahlström A, Sayin SI, Marschall HU, Bäckhed F. Intestinal crosstalk between bile acids and microbiota and its impact on host metabolism. *Cell Metab.* 2016;24(1):41–50. doi:10.1016/j.cmet.2016.05.005. PMID: 27320064.
138. Grüner N, Mattner J. Bile acids and microbiota: multifaceted and versatile regulators of the liver–gut axis. *Int J Mol Sci.* 2021;22(3):1397. doi:10.3390/ijms22031397. PMID: 33573273.
139. Basuray N, Deehan EC, Vieira FT, Avedzi HM, Duke RL, Colín-Ramírez E, Tun HM, Zhang Z, Wine E, Madsen KL, et al. Dichotomous effect of dietary fiber in pediatrics: a narrative review of the health benefits and tolerance of fiber. *Eur J Clin Nutr.* 2024;78(7):557–568. doi:10.1038/s41430-024-01429-5. PMID: 38480843.
140. Kelishadi R, Farajian S, Safavi M, Mirlohi M, Hashemipour M. A randomized triple-masked controlled trial on the effects of synbiotics on inflammation markers in overweight children. *J Pediatr (Rio J).* 2014;90(2):161–168. doi:10.1016/j.jpmed.2013.07.003. PMID: 24184299.
141. Jones RB, Alderete TL, Kim JS, Millstein J, Gilliland FD, Goran MI. High intake of dietary fructose in overweight/obese teenagers associated with depletion of Eubacterium and streptococcus in gut microbiome. *Gut Microbes.* 2019;10(6):712–719. doi:10.1080/19490976.2019.1592420. PMID: 30991877.
142. Rampelli S, Guenther K, Turroni S, Wolters M, Veidebaum T, Kourides Y, Molnár D, Lissner L, Benitez-Paez A, Sanz Y, et al. Pre-obese children's dysbiotic gut microbiome and unhealthy diets may predict the development of obesity. *Commun Biol.* 2018;1(1):222. doi:10.1038/s42003-018-0221-5. PMID: 30534614.
143. Leong C, Haszard JJ, Heath AM, Tannock GW, Lawley B, Cameron SL, Szymlek-Gay EA, Gray AR, Taylor BJ, Galland BC, et al. Using compositional principal component analysis to describe children's gut microbiota in relation to diet and body composition. *Am J Clin Nutr.* 2020;111(1):70–78. doi:10.1093/ajcn/nqz270. PMID: 31711093.
144. Kisuse J, La-Ongkham O, Nakphaichit M, Therdtatha P, Momoda R, Tanaka M, Fukuda S, Popluechai S, Kespechara K, Sonomoto K, et al. Urban diets linked to gut microbiome and metabolome alterations in children: a comparative cross-sectional study in Thailand. *Front Microbiol.* 2018;9:1345. doi:10.3389/fmicb.2018.01345. PMID: 29988433.
145. Li H, Zhao L, Zhang M. Gut microbial SNPs induced by high-fiber diet dominate nutrition metabolism and environmental adaptation of faecalibacterium prausnitzii in obese children. *Front Microbiol.* 2021;12:683714. doi:10.3389/fmicb.2021.683714. PMID: 34135881.
146. Zhang C, Yin A, Li H, Wang R, Wu G, Shen J, Zhang M, Wang L, Hou Y, Ouyang H, et al. Dietary modulation of gut microbiota contributes to alleviation of both genetic and simple obesity in children. *EBioMedicine.* 2015;2(8):968–984. doi:10.1016/j.ebiom.2015.07.007. PMID: 26425705.
147. Orbe-Orihuela YC, Godoy-Lozano EE, Lagunas-Martínez A, Castañeda-Márquez AC, Murga-Garrido S, Díaz-Benítez CE, Ochoa-Leyva A, Cornejo-Granados F, Cruz M, Estrada K, et al. Association of gut microbiota with dietary-dependent childhood obesity. *Arch Med Res.* 2022;53(4):407–415. doi:10.1016/j.arcmed.2022.03.007. PMID: 35382951.
148. Kilic Yildirim G, Dinleyici M, Vandenplas Y, Dinleyici EC. Effects of synbiotic supplementation on intestinal microbiota composition in children and adolescents with exogenous obesity: (probesity-2 trial). *Gut Pathog.* 2023;15(1):36. doi:10.1186/s13099-023-00563-y.
149. Nicolucci AC, Hume MP, Martínez I, Mayengbam S, Walter J, Reimer RA. Prebiotics reduce body fat and alter intestinal microbiota in children who are overweight or with obesity. *Gastroenterology.* 2017;153(3):711–722. doi:10.1053/j.gastro.2017.05.055. PMID: 28596023.

150. Wu GD, Chen J, Hoffmann C, Bittinger K, Chen YY, Keilbaugh SA, Bewtra M, Knights D, Walters WA, Knight R, et al. Linking long-term dietary patterns with gut microbial enterotypes. *Science*. 2011;334(6052):105–108. doi:10.1126/science.1208344. PMID: 21885731.
151. Flint HJ, Scott KP, Louis P, Duncan SH. The role of the gut microbiota in nutrition and health. *Nat Rev Gastroenterol Hepatol*. 2012;9(10):577–589. doi:10.1038/nrgastro.2012.156. PMID: 22945443.
152. Flint HJ, Scott KP, Duncan SH, Louis P, Forano E. Microbial degradation of complex carbohydrates in the gut. *Gut Microbes*. 2012;3(4):289–306. doi:10.4161/gmic.19897. PMID: 22572875.
153. Garcia-Ribera S, Amat-Bou M, Climent E, Llobet M, Chenoll E, Corripio R, Ibáñez L, Ramon-Krauel M, Lerin C. Specific dietary components and gut microbiota composition are associated with obesity in children and adolescents with prader-Willi Syndrome. *Nutrients*. 2020;12(4):12. doi:10.3390/nu12041063. PMID: 32290434.
154. Qin W, Sun L, Miao M, Zhang G. Plant-sourced intrinsic dietary fiber: physical structure and health function. *Trends Food Sci Technol*. 2021;118:341–355. doi:10.1016/j.tifs.2021.09.022.
155. Koh A, De Vadder F, Kovatcheva-Datchary P, Bäckhed F. From dietary fiber to host physiology: short-chain fatty acids as key bacterial metabolites. *Cell*. 2016;165(6):1332–1345. doi:10.1016/j.cell.2016.05.041. PMID: 27259147.
156. Ferro LE, Crowley LN, Bittinger K, Friedman ES, Decker JE, Russel K, Katz S, Kim JK, Trabulsi JC. Effects of prebiotics, probiotics, and synbiotics on the infant gut microbiota and other health outcomes: a systematic review. *Crit Rev Food Sci Nutr*. 2023;63(22):5620–5642. doi:10.1080/10408398.2021.2022595. PMID: 37667870.
157. Walter J, Maldonado-Gómez MX, Martínez I. To engraft or not to engraft: an ecological framework for gut microbiome modulation with live microbes. *Curr Opin Biotechnol*. 2018;49:129–139. doi:10.1016/j.copbio.2017.08.008. PMID: 28866242.
158. Reynés B, Palou M, Rodríguez AM, Palou A. Regulation of adaptive thermogenesis and Browning by prebiotics and postbiotics. *Front Physiol*. 2018;9:1908. doi:10.3389/fphys.2018.01908. PMID: 30687123.
159. Duque A, Demarqui FM, Santoni MM, Zanelli CF, Adorno MAT, Milenkovic D, Mesa V, Sivieri K. Effect of probiotic, prebiotic, and synbiotic on the gut microbiota of autistic children using an in vitro gut microbiome model. *Food Res Int*. 2021;149:110657. doi:10.1016/j.foodres.2021.110657. PMID: 34600659.
160. Franssen F, Sahasrabudhe NM, Elderman M, Bosveld M, El Aidy S, Hugenholtz F, Borghuis T, Kousemaker B, Winkel S, van der Gaast-de Jongh C, et al.  $\beta$ 2 $\rightarrow$ 1-fructans modulate the immune system in vivo in a microbiota-dependent and -Independent fashion. *Front Immunol*. 2017;8:154. doi:10.3389/fimmu.2017.00154. PMID: 28261212.
161. Lombard V, Golaconda Ramulu H, Drula E, Coutinho PM, Henrissat B. The carbohydrate-active enzymes database (CAZy) in 2013. *Nucl Acids Res*. 2014;42(D1):D490–5. doi:10.1093/nar/gkt1178. PMID: 24270786.
162. El Kaoutari A, Armougom F, Gordon JI, Raoult D, Henrissat B. The abundance and variety of carbohydrate-active enzymes in the human gut microbiota. *Nat Rev Microbiol*. 2013;11(7):497–504. doi:10.1038/nrmicro3050. PMID: 23748339.
163. Fulgoni VL 3rd, Brauchla M, Fleige L, Chu Y. Association of whole-grain and dietary fiber intake with cardiometabolic risk in children and adolescents. *Nutr Health*. 2020;26(3):243–251. doi:10.1177/0260106020928664. PMID: 32552292.
164. Hume MP, Nicolucci AC, Reimer RA. Prebiotic supplementation improves appetite control in children with overweight and obesity: a randomized controlled trial. *Am J Clin Nutr*. 2017;105(4):790–799. doi:10.3945/ajcn.116.140947. PMID: 28228425.
165. Barczynska R, Slizewska K, Litwin M, Szalecki M, Kapusniak J. Effects of dietary fiber preparations made from maize starch on the growth and activity of selected bacteria from the Firmicutes, Bacteroidetes, and Actinobacteria phyla in fecal samples from obese children. *Acta Biochim Pol*. 2016;63(2):261–266. doi:10.18388/abp.2015\_1068. PMID: 26929930.
166. Cani PD, de Vos WM. Next-generation beneficial microbes: the case of *Akkermansia muciniphila*. *Front Microbiol*. 2017;8:1765. doi:10.3389/fmicb.2017.01765. PMID: 29018410.
167. Everard A, Belzer C, Geurts L, Ouwerkerk JP, Druart C, Bindels LB, Guiot Y, Derrien M, Muccioli GG, Delzenne NM, et al. Cross-talk between *Akkermansia muciniphila* and intestinal epithelium controls diet-induced obesity. *Proc Natl Acad Sci USA*. 2013;110(22):9066–9071. doi:10.1073/pnas.1219451110. PMID: 23671105.
168. Lam KC, Araya RE, Huang A, Chen Q, Di Modica M, Rodrigues RR, Lopès A, Johnson SB, Schwarz B, Bohrnsen E, et al. Microbiota triggers sting-type I ifn-dependent monocyte reprogramming of the tumor microenvironment. *Cell*. 2021;184(21):5338–56.e21. doi:10.1016/j.cell.2021.09.019. PMID: 34624222.
169. Schmidt-Wittig U, Enss ML, Coenen M, Gärtner K, Hedrich HJ. Response of rat colonic mucosa to a high fiber diet. *Ann Nutr Metab*. 1996;40(6):343–350. doi:10.1159/000177936. PMID: 9087313.
170. Kim Y, Hwang SW, Kim S, Lee YS, Kim TY, Lee SH, Kim SJ, Yoo HJ, Kim EN, Kweon MN. Dietary cellulose prevents gut inflammation by modulating lipid metabolism and gut microbiota. *Gut Microbes*. 2020;11(4):944–961. doi:10.1080/19490976.2020.1730149. PMID: 32138587.

171. Procházková N, Falony G, Dragsted LO, Licht TR, Raes J, Roager HM. Advancing human gut microbiota research by considering gut transit time. *Gut*. 2023;72(1):180–191. doi:10.1136/gutjnl-2022-328166. PMID: 36171079.
172. Deehan EC, Yang C, Perez-Muñoz ME, Nguyen NK, Cheng CC, Triador L, Zhang Z, Bakal JA, Walter J. Precision microbiome modulation with discrete dietary fiber structures directs short-chain fatty acid production. *Cell Host Microbe*. 2020;27(3):389–404. e6. doi:10.1016/j.chom.2020.01.006.
173. Puhlmann ML, de Vos WM. Intrinsic dietary fibers and the gut microbiome: rediscovering the benefits of the plant cell matrix for human health. *Front Immunol*. 2022;13:954845. doi:10.3389/fimmu.2022.954845. PMID: 36059540.
174. Dietary Reference Intakes: Nutrient recommendations and databases [Internet]. National Academies Press; 2005 [cited 2023 Apr]. <https://ods.od.nih.gov/HealthInformation/nutrientrecommendations.aspx>.
175. Deehan EC, Mocanu V, Madsen KL. Effects of dietary fibre on metabolic health and obesity. *Nat Rev Gastroenterol Hepatol*. 2024;21(5):301–318. doi:10.1038/s41575-023-00891-z. PMID: 38326443.
176. McRorie JW Jr., McKeown NM. Understanding the physics of functional fibers in the gastrointestinal tract: an evidence-based approach to resolving enduring misconceptions about insoluble and soluble fiber. *J Acad Nutr Diet*. 2017;117(2):251–264. doi:10.1016/j.jand.2016.09.021. PMID: 27863994.
177. Hoek M, Merks RMH. Emergence of microbial diversity due to cross-feeding interactions in a spatial model of gut microbial metabolism. *BMC Syst Biol*. 2017;11(1):56. doi:10.1186/s12918-017-0430-4. PMID: 28511646.
178. Moens F, Weckx S, De Vuyst L. Bifidobacterial inulin-type fructan degradation capacity determines cross-feeding interactions between bifidobacteria and faecalibacterium prausnitzii. *Int J Food Microbiol*. 2016;231:76–85. doi:10.1016/j.ijfoodmicro.2016.05.015. PMID: 27233082.
179. Mills RH, Dulai PS, Vázquez-Baeza Y, Saucedo C, Daniel N, Gerner RR, Batachari LE, Malfavon M, Zhu Q, Weldon K, et al. Multi-omics analyses of the ulcerative colitis gut microbiome link bacteroides vulgatus proteases with disease severity. *Nat Microbiol*. 2022;7(2):262–276. doi:10.1038/s41564-021-01050-3. PMID: 35087228.
180. Mars RAT, Yang Y, Ward T, Houtti M, Priya S, Lekatz HR, Tang X, Sun Z, Kalari KR, Korem T, et al. Longitudinal multi-omics reveals subset-specific mechanisms underlying irritable bowel syndrome. *Cell*. 2020;182(6):1460–73.e17. doi:10.1016/j.cell.2020.08.007. PMID: 32916129.
181. de Cuevillas B, Milagro FI, Tur JA, Gil-Campos M, de Miguel-Etayo P, Martínez JA, Navas-Carretero S. Fecal microbiota relationships with childhood obesity: a scoping comprehensive review. *Obes Rev*. 2022;23(Suppl 1):e13394. doi:10.1111/obr.13394. PMID: 34913242.
182. Liber A, Szajewska H. Effect of oligofructose supplementation on body weight in overweight and obese children: a randomised, double-blind, placebo-controlled trial. *Br J Nutr*. 2014;112(12):2068–2074. doi:10.1017/s0007114514003110. PMID: 25327394.
183. Reynolds AN, Akerman AP, Mann J. Dietary fibre and whole grains in diabetes management: systematic review and meta-analyses. *PLoS Med*. 2020;17(3):e1003053. doi:10.1371/journal.pmed.1003053. PMID: 32142510.
184. Deehan EC, Duar RM, Armet AM, Perez-Muñoz ME, Jin M, Walter J, Britton RA, Cani PD. Modulation of the gastrointestinal microbiome with nondigestible fermentable carbohydrates to improve human health. *Microbiol Spectr*. 2017;5(5). doi:10.1128/microbiol.spec.BAD-0019-2017. PMID: 28936943.
185. Lichtenstein AH, Petersen K, Barger K, Hansen KE, Anderson CAM, Baer DJ, Lampe JW, Rasmussen H, Matthan NR. Perspective: design and conduct of human nutrition randomized controlled trials. *Adv Nutr*. 2021;12(1):4–20. doi:10.1093/advances/nmaa109. PMID: 33200182.
186. Kane PB, Bittlinger M, Kimmelman J. Individualized therapy trials: navigating patient care, research goals and ethics. *Nat Med*. 2021;27(10):1679–1686. doi:10.1038/s41591-021-01519-y. PMID: 34642487.
187. Ghosh TS, Das M, Jeffery IB, O'Toole PW. Adjusting for age improves identification of gut microbiome alterations in multiple diseases. *Elife*. 2020;9. doi:10.7554/eLife.50240. PMID: 32159510.
188. de la Cuesta-Zuluaga J, Kelley ST, Chen Y, Escobar JS, Mueller NT, Ley RE, McDonald D, Huang S, Swafford AD, Knight R, et al. Age- and sex-dependent patterns of gut microbial diversity in human adults. *mSystems*. 2019;4(4). doi:10.1128/mSystems.00261-19. PMID: 31098397.
189. Bowyer RCE, Jackson MA, Pallister T, Skinner J, Spector TD, Welch AA, Steves CJ. Use of dietary indices to control for diet in human gut microbiota studies. *Microbiome*. 2018;6(1):77. doi:10.1186/s40168-018-0455-y. PMID: 29695307.
190. Meslier V, Laiola M, Roager HM, De Filippis F, Roume H, Quinquis B, Giacco R, Mennella I, Ferracane R, Pons N, et al. Mediterranean diet intervention in overweight and obese subjects lowers plasma cholesterol and causes changes in the gut microbiome and metabolome independently of energy intake. *Gut*. 2020;69(7):1258–1268. doi:10.1136/gutjnl-2019-320438. PMID: 32075887.
191. Nguyen NK, Deehan EC, Zhang Z, Jin M, Baskota N, Perez-Muñoz ME, Cole J, Tuncil YE, Seethaler B, Wang T, et al. Gut microbiota modulation with long-chain corn bran arabinoxylan in adults with

- overweight and obesity is linked to an individualized temporal increase in fecal propionate. *Microbiome*. 2020;8(1):118. doi:10.1186/s40168-020-00887-w. PMID: 32814582.
192. Walter J, Armet AM, Finlay BB, Shanahan F. Establishing or exaggerating causality for the gut microbiome: lessons from human microbiota-associated rodents. *Cell*. 2020;180:221–232. doi:10.1016/j.cell.2019.12.025. PMID: 31978342.
193. Schmidt F, Zimmermann J, Tanna T, Farouni R, Conway T, Macpherson AJ, Platt RJ. Noninvasive assessment of gut function using transcriptional recording sentinel cells. *Science*. 2022;376(6594): eabm6038. doi:10.1126/science.abm6038. PMID: 35549411.
194. Arrieta MC, Walter J, Finlay BB. Human microbiota-associated mice: a model with challenges. *Cell Host Microbe*. 2016;19(5):575–578. doi:10.1016/j.chom.2016.04.014. PMID: 27173924.
195. Paul W, Marta C, Tom VW. Resolving host-microbe interactions in the gut: the promise of in vitro models to complement in vivo research. *Curr Opin Microbiol*. 2018;44:28–33. doi:10.1016/j.mib.2018.07.001. PMID: 30031243.
196. Deehan EC, Zhang Z, Riva A, Armet AM, Perez-Muñoz ME, Nguyen NK, Krysa JA, Seethaler B, Zhao YY, Cole J, et al. Elucidating the role of the gut microbiota in the physiological effects of dietary fiber. *Microbiome*. 2022;10(1):77. doi:10.1186/s40168-022-01248-5. PMID: 35562794.